





Thermal immuno-nanomedicine in cancer

Zhe Yang^{1,7} , Di Gao^{1,7}, Jing Zhao^{2,7}, Gaojie Yang³, Ming Guo³, Ying Wang¹, Xuechun Ren¹, Jong Seung Kim⁴, Lin Jin⁵ , Zhongmin Tian¹  & Xingcai Zhang⁶ 

Abstract

Immunotherapy has revolutionized the treatment of patients with cancer. However, promoting antitumour immunity in patients with tumours that are resistant to these therapies remains a challenge. Thermal therapies provide a promising immune-adjuvant strategy for use with immunotherapy, mostly owing to the capacity to reprogramme the tumour microenvironment through induction of immunogenic cell death, which also promotes the recruitment of endogenous immune cells. Thus, thermal immunotherapeutic strategies for various cancers are an area of considerable research interest. In this Review, we describe the role of the various thermal therapies and provide an update on attempts to combine these with immunotherapies in clinical trials. We also provide an overview of the preclinical development of various thermal immuno-nanomedicines, which are capable of combining thermal therapies with various immunotherapy strategies in a single therapeutic platform. Finally, we discuss the challenges associated with the clinical translation of thermal immuno-nanomedicines and emphasize the importance of multidisciplinary and inter-professional collaboration to facilitate the optimal translation of this technology from bench to bedside.

Sections

[Introduction](#)[Thermal therapies](#)[Combinations with immunotherapy](#)[Thermal immuno-nanomedicines](#)[Future prospects and challenges](#)[Conclusions](#)

¹The Key Laboratory of Biomedical Information Engineering of Ministry of Education, School of Life Science and Technology, Xi'an Jiaotong University, Xi'an, China. ²Shaanxi Key Laboratory of Degradable Biomedical Materials, School of Chemical Engineering, Northwest University, Xi'an, China. ³School of Engineering, Massachusetts Institute of Technology, Cambridge, MA, USA. ⁴Department of Chemistry, Korea University, Seoul, Republic of Korea. ⁵International Joint Research Laboratory for Biomedical Nanomaterials of Henan, Zhoukou Normal University, Zhoukou, China. ⁶John A. Paulson School of Engineering and Applied Sciences, Harvard University, Cambridge, MA, USA. ⁷These authors contributed equally: Zhe Yang, Di Gao, Jing Zhao. ✉ e-mail: yangzhe@xjtu.edu.cn; jinlin_1982@126.com; zmtian@xjtu.edu.cn; chemmike1984@gmail.com

Key points

- Immunotherapy has revolutionized cancer therapy, and the clinical effectiveness of approaches such as immune-checkpoint inhibitors and cellular immunotherapies has created substantial improvements in outcomes.
- Thermal therapies designed to deliver local hyperthermia can promote antitumour immunity via the induction of immunogenic cell death following tumour ablation and by reprogramming the tumour microenvironment.
- Several early-phase trials combining thermal therapies with immunotherapy are either ongoing or completed, and some have provided encouraging results that support the further clinical development of combined thermal therapy–immunotherapy approaches.
- Thermal immuno-nanomedicines are thermal therapies that also incorporate immunotherapies within a single nanoparticle, potentially enabling simultaneous activity and synergy between these two modalities.
- Preclinical evidence suggests that thermal immuno-nanomedicines provide a promising cancer therapeutic strategy; however, coordinated efforts from multidisciplinary teams of experts will be required to drive the clinical translation of this technology.

Introduction

Immunotherapy has revolutionized cancer therapy. Several clinically effective cancer immunotherapy strategies have been established, including immune-checkpoint inhibition, vaccination and adoptive T cell transfer, comprising chimeric antigen receptor (CAR)-engineered and T cell receptor (TCR)-engineered T cell therapies^{1–6}. However, cancers have a notorious ability to evade the immune system, and tumours that are not inflamed and/or that do not elicit an immune response, so-called ‘cold’ tumours, often do not respond to immunotherapies^{7–10}. Furthermore, the immunosuppressive tumour microenvironment (TME) often acts as a barrier to immune cell infiltration that enables the tumour to evade immune surveillance, leading to tumour progression^{11–13}. Accordingly, methods of reprogramming the TME to convert cold tumours to hot tumours and thus improve the efficacy of immunotherapy are a focus of considerable research interest.

To address these challenges, strategies combining immunotherapy with other therapeutic modalities have been developed and tested both in preclinical models and in clinical trials^{14–20}. For example, more than eight regimens combining immune-checkpoint inhibitors (ICIs) with chemotherapy have been approved by the FDA for patients with metastatic non-small-cell lung cancer (NSCLC) and the combination of ICIs with radiotherapy is being explored for the same indication in several trials (including NCT03867175, NCT03774732, NCT03446911)^{21–23}. Thermal therapy, which involves heating either the whole body or local areas, such as the tumour site, is a field of increasing research interest, mostly owing to the capacity to improve clinical responses to radiotherapy, chemotherapy and immunotherapy in patients with, among others, breast cancer, colon cancer or central nervous system cancers^{24–29}. Thus, the FDA has approved several devices designed to

deliver thermal therapies, including high-intensity focused ultrasound (HIFU) and radiofrequency ablation (RFA), for hepatocellular carcinomas (HCCs), liver metastases and renal cell carcinomas as well as other cancers^{30,31} (Tables 1 and 2).

Thermal therapy has been shown to induce immunogenic cell death (ICD) of cancer cells, which promotes both innate and adaptive immunity in preclinical models^{32–34} (Fig. 1). Moreover, thermal therapy improves perfusion and reduces intratumour interstitial pressure, most likely owing to a transient increase in vascular permeability²⁶, which promotes the accumulation of therapeutic co-stimulatory molecules and/or immune cells and facilitates the release of pro-inflammatory cytokines and chemokines that can overcome an immunosuppressive TME^{35–38}. Various clinical trials combining thermal therapy and immunotherapy have been conducted and could lead to advances in cancer therapy^{39–41}.

Despite the encouraging performance of thermal immunotherapy in preclinical models^{41,42}, several limitations exist that are likely to impair the therapeutic effectiveness of this approach in patients with cancer. Firstly, the usually systemic administration and non-specific accumulation of immunotherapeutic and/or thermotherapeutic agents not only influence their antitumour activities but also can result in adverse events⁴³; secondly, in the future, gene-editing technologies are increasingly likely to be employed to modify tumour cells and/or their microenvironment, including regulating the expression of immune checkpoints and, potentially, of other targets of interest, and this will require safe and effective transfection systems^{44–46}. Data from an increasing number of studies demonstrate that nanomedicines have considerable potential to overcome these challenges owing to several unique advantages over traditional drug formulations, such as enhanced targeting capacity, intracellular delivery, controlled release of therapeutic agents, bioavailability and multi-functionality. Thus, thermal immuno-nanomedicines (TINs), which involve the nanoparticle-mediated delivery of thermal therapies alongside an immunotherapeutic agent that enables simultaneous synergy with thermal therapy, are an area of rapidly developing research interest^{34,47,48}.

In this Review, we summarize the current status and development of thermal therapies and attempt to combine these therapies with immunotherapies. We provide an overview of the various TIN strategies, which have the potential to deliver various thermal therapies combined with other cutting-edge immunotherapy approaches. We also highlight the challenges that are likely to arise in the clinical translation of TINs and emphasize the importance of coordinated multidisciplinary teams comprising experts with various professional backgrounds to optimize clinical translation.

Thermal therapies

The activity of biological systems is profoundly affected by even small elevations in temperature. A well-controlled increase in temperature can support the treatment of various diseases, including cancer^{49–51}. Over the past few decades, substantial progress has been made in developing different heating techniques with improved temperature control that maximize therapeutic efficacy while minimizing the risk of adverse events. According to the extent of the temperature increment, therapeutic heating modalities can be broadly categorized as hyperthermia or thermal ablation.

Hyperthermia refers to mild, transient elevations in tissue temperature to a maximum of 45 °C (ref. 52). Clinically, regional hyperthermia can improve the therapeutic effect of more traditional therapies through various mechanisms such as the promotion of blood flow

Table 1 | Commonly used electromagnetic thermal therapeutic devices for cancer therapy

Device	Cancer type	Main features
RFA		
Cool-tip RFA system (Valleylab/Radionics/Covidien, USA) ²⁵⁶	Unresectable liver lesions (primary or metastatic) ⁷¹	Monopolar equipment; straight-needle design; maximum energy delivery (E series)
RITA StarBurst RFA system (AngioDynamics, USA) ²⁵⁷	Primary liver tumours ⁷² , metastases from primary CRCs ⁷⁵	Multipolar equipment; real-time tissue monitoring at the margins of the ablation
RF 3000 RFA system with LeVein needle electrodes (Boston Scientific, USA) ²⁵⁸	Primary HCCs ⁷³	Multipolar equipment; wide portfolio matrix of array diameters from 2 to 5 cm; soloist single-needle electrode for small 1.5 cm × 1 cm ablation zones
Celon Olympus (Medical and Industrial Equipment, UK) ²⁵⁹	HCCs ⁷⁴ , RCCs ⁷⁶ , osteoid osteomas ⁸¹	Integration of bipolar and multipolar probes into a single equipment; intratumoural and extratumoural (no-touch technique) placement of applicators; large range of ablation diameters (5–90 mm); optimized treatment by resistance-controlled automatic power
HALO ³⁶⁰⁺ bipolar RFA system (Covidien GI Solutions, USA) ²⁶⁰	Barrett oesophagus ⁷⁷	Automated radiofrequency energy delivery; fixed amount of radiofrequency energy density; fixed power; bipolar electrode array
HALO ⁹⁰ RFA system (Covidien GI Solutions, USA) ²⁶¹	Barrett oesophagus ⁷⁸	Used for secondary RFA of residual oesophageal lesions after initial circumferential ablation using the HALO ³⁶⁰⁺ system
OsteoCool RFA system (Medtronic, USA) ²⁶²	Spinal metastases ⁸⁰	Coaxial and bipolar technology delivers RFA, cooled with circulating water
OptaBlate Bone Tumour Ablation System (Stryker, USA) ²⁶³	Bone tumours	Microinfusion technology enabling the concurrent treatment of two vertebral body levels using a bipedicular approach
S-5L RFA system (MedSphere International, China) ⁷⁹	Mixed thyroid nodules ⁷⁹	Simultaneously compatible with internally cooled and normal system; dual-mode control-power control and temperature control; real-time temperature, power and resistance feedback
MWA		
Emprint ablation system (Medtronic, USA) ²⁶⁴	Primary HCCs or liver metastases ⁸⁷	Thermosphere technology to control the microwave field and length of the microwaves; more spherical and more predictable ablations
NeuWave Certus MWA system (NeuWave Medical, USA) ²⁶⁵	Primary HCCs or liver metastases ⁸⁸	Probe–multiprobe synchrony helps to protect non-target tissues; versatile probe portfolio to control the ablation size, shape and position; reduced blood loss, procedure time and length of hospital stay with pre-transection coagulation
HS AMICA (HS Hospital Service, Italy) ²⁶⁶	HCCs ⁹⁰ , bone metastases ⁹³	Capable of both microwave and radiofrequency energy generation, delivery and monitoring; embedded peristaltic pump enables internal cooling of either type of applicator
Acculis MWA system (AngioDynamics, USA) ²⁶⁷	NSCLCs ⁹² , HCCs ⁹¹	The fastest device in achieving the largest coagulation diameter (short-axis) at 5 and 6 min; predictable ablation zones
Evident MWA System (Covidien, USA) ²⁶⁸	HCCs, CRCs ⁶¹	Minimized set-up, simultaneous antenna use, multiple-use antennas
LITT		
Visualase Thermal Therapy System (Medtronic, USA) ²⁶⁹	Meningiomas, ependymoma, gliomas ⁸²	980-nm laser with power of 15 W; capable of MRI-guided laser ablation; smallest laser catheter (1.65 mm in diameter) on the market; requires minimal sutures, typically a single stitch; requires no ionizing radiation or large skull flap
NeuroBlate System (Monteris Medical, Canada) ²⁷⁰	Meningiomas ⁶³ , gliomas ⁶⁴	1,064-nm laser with power of 12 W; MRI-guided laser ablation; robotically controlled probe driver; mini-bolt access; side-firing probes for treatment of irregularly shaped lesions

CRC, colorectal cancer; HCC, hepatocellular carcinoma; LITT, laser interstitial thermal therapy; MWA, microwave ablation; NSCLC, non-small-cell lung cancer; RCC, renal cell carcinoma; RFA, radiofrequency ablation.

resulting in tumour reoxygenation (39–44 °C), prevention of DNA damage repair (>41 °C) and activation of an antitumour immune response^{53,54}. Thermal ablation refers to the application of high temperatures (>50 °C) to directly damage tumour cells via coagulation and protein denaturation. Clinically, thermal ablation has been recognized as a minimally invasive alternative to certain surgical procedures such as ablation of early-stage HCCs⁵⁵. Owing to differences in energy transduction and delivery techniques between high-grade heating devices, the heating depth, direction and volume can all vary between devices, each with its own advantages and disadvantages⁵⁰. In this section, common thermal therapeutic modalities and their applications in the clinic are described in detail (Tables 1 and 2 and Fig. 2).

Electromagnetic thermal therapy

Electromagnetic thermal therapy harnesses the thermal energy produced by a high-frequency alternating sinusoidal electromagnetic field. Two main mechanisms of heat generation are involved in electromagnetic heating: dielectric heating and ionic conduction. Polar molecules, such as water, preferentially align in succession in the alternating electromagnetic field owing to their inherent electric dipole moment upon exposure to an alternating current. As a consequence, these molecules acutely rotate with frequencies in the MHz range and the obtained excessive kinetic energy then dissipates owing to collisions with other molecules, leading to so-called dielectric heating³⁶. Ions located in tissues generally oscillate in response to

electromagnetic stimulation in the kilohertz range⁵⁷. Owing to an abundance of neighbouring molecules and atoms, the oscillation of ions and the resultant dispersal of kinetic energy form an effective method of generating heat⁵⁷.

Three main electromagnetic heating-based thermal therapy modalities – RFA, microwave ablation (MWA) and photothermal therapy (PTT) – are currently being either applied in clinical practice or investigated in clinical trials^{51,58–64}. The clinical applications of PTT, including laser interstitial thermal therapy, have been comprehensively reviewed elsewhere⁶⁵; therefore, this discussion is focused on RFA and MWA. Several commercially available interstitial heating devices are currently available. For most devices used during both RFA and MWA, a needle-shaped catheter is surgically implanted into the targeted tissues, often with guidance by medical imaging, and heat is then generated locally via the catheter, known as interstitial heating.

Radiofrequency ablation. RFA is currently the most widely used clinical electromagnetic ablative modality and has the advantages of minimal invasiveness and a limited risk of adverse events, making this approach a reasonable option when surgical resection is inadvisable^{66–69}. To deliver RFA, a slender probe is inserted into the tumour tissues with imaging guidance, followed by the creation of an alternating current (350–500 kHz) within the target tissues. The probes, also known as RFA electrodes, can be divided into single-tip monopolar probes and multipolar cluster arrays with the former requiring grounding pads to close the electric circuit. To achieve the desired anchoring and ablation profiles, umbrella-shaped clustered electrodes have also been developed. In this scenario, a carefully controlled amount of energy is delivered to the tumour through the electrodes to induce a uniform heating zone in direct proximity to the probe⁷⁰. The electrical conductivity of the target tissue is usually enhanced, for example, using intra-tumoural saline injections, to deliver a sufficiently high temperature to cover the entire target tissue. Some other functional probes, such as multitoned expandable probes, needle perfusion probes and cool-tip probes, are also available for different application scenarios⁵⁷. Several RFA devices have been approved by the FDA for clinical application in the USA, including the Cool-Tip RFA system, the OsteoCool RF ablation system and the OptaBlate ablation system (Table 1). Clinical indications for RFA include several thoracic and gastrointestinal tumours, including liver lesions^{71–74}, colorectal cancers⁷⁵, renal cell carcinomas⁷⁶, oesophageal cancers^{77,78}, thyroid nodules⁷⁹, spinal metastases⁸⁰ and osteoid osteomas⁸¹. An example of the effectiveness of RFA is provided by a multicentre, open-label trial in which 90 patients with liver cancer with a Child–Pugh classification of A or B and either multiple (≤ 3 in number and ≤ 3 cm in diameter) or solitary (≤ 4 cm in diameter) lesions underwent bipolar RFA using the Celon Olympus RFA system. Complete tumour necrosis was achieved in 97.8% of patients. Three patients had adverse events requiring hospitalization (abdominal wall burn, pleural effusion and biliary peritonitis)⁸².

Microwave ablation. MWA provides another method of inducing immediate coagulation necrosis of the target lesion as seen with RFA. During MWA, microwaves with specific frequencies (typically 915–2,450 MHz) are generated using a magnetron in a microwave therapeutic apparatus and delivered to tumour tissues through an intratumourally placed needle-shaped antenna. Similar to RFA, dielectric heating from the inherent dipoles is responsible for thermogenesis for MWA⁸³. However, unlike RFA, MWA does not require an electric current, which alleviates the concerns regarding tissue desiccation

Table 2 | Commonly used HIFU systems for cancer therapy

Device	Cancer type	Main features
Sonablate system (InnoMedicus, Austria) ²⁷¹	Prostate cancer ⁹⁹	Transrectal HIFU system; targets specific tissues using ultrasound MR fusion; enables ablation with pinpoint accuracy while sparing untargeted tissues; analysis using real-time imaging and advanced tissue change monitoring technology
Ablatherm system (EDAP TMS, USA) ²⁷²	Prostate cancer ¹⁰⁰	Transrectal HIFU system; MR images and biopsy maps imported for automatic matching of ultrasound images; robotic procedure; preservation of surrounding tissue with different treatment strategies (whole gland/nerve sparing/partial gland)
TULSA-PRO system (Profound, USA) ²⁷³	Prostate cancer ¹⁰¹	Transurethral HIFU system; MRI-guided device positioning; robotically driven ultrasound ablation with closed-loop thermal feedback control; capable of sweeping ultrasound and continuous rotation; capable of treating both large and small prostate volumes, anterior and posterior prostate tissues; enables thermal protection of urethra and rectum
ExAblate system (Insightec, Israel) ²⁷⁴	Prostate cancer ⁹⁶	Enables real-time therapeutic monitoring
Sonalleve system (Profound, USA) ²⁷⁵	Desmoid tumours ¹⁰⁶ , breast cancer ¹⁰⁵ , osteoid osteomas ¹⁰³	Enables real-time MRI temperature monitoring and control; reduced cooling time between sonications with direct skin cooling and dual-mode thermometry
Haifu Model JC (Chongqing Haifu Medical Technology, China) ²⁷⁶	Primary HCCs or metastases, pancreatic cancers ¹⁰⁷ , RCCs ¹⁰⁹	Enables conformal and precise ablation; one-off treatment, not limited by tumour size and shape; real-time ultrasound-guided therapy with digital quantitative analysis
HIFUNIT9000 (Shanghai A&S Science Technology Development, China) ²⁷⁷	Pancreatic cancers ¹⁰⁸	Real-time ultrasound-guided therapy; enables intelligent three-dimensional reconstruction of tumours; multiple arrays and double focuses

HCC, hepatocellular carcinoma; HIFU, high-intensity focused ultrasound; RCC, renal cell carcinoma.

during the therapeutic process, which can compromise the therapeutic effectiveness of RFA. This important difference in heat source makes MWA more appropriate for lesions located in tissues with higher impedance and limited thermal conduction such as lung and bone⁸⁴. High-frequency MWA ($\geq 2,450$ MHz) typically generates an ablation area that is smaller and rounder than that obtained with low-frequency MWA, making this method more suitable for the ablation of small, round tumours, while avoiding damage to surrounding non-malignant tissues. Thus, predicting the size of the ablation zone becomes easier with high-frequency MWA. However, lower-frequency MWA has the advantage of deeper tissue penetration and higher energy conversion

efficiency, which are both beneficial when treating larger tumours^{85,86}. Thus far, two MWA devices, Emprint and Evident MWA, have received FDA approval for the treatment of patients with liver tumours, including primary HCCs and liver metastases (Table 1). To evaluate the efficacy of Emprint MWA for the ablation of unresectable large-diameter liver lesions (≥ 3 cm in diameter), 21 patients with such tumours (mean diameter of 34.7 mm) underwent percutaneous MWA. The shape and volume of the ablation zones were evaluated using CT imaging within the month before and the month after treatment. Residual tumour material was observed in only 4.8% of tumours after a single ablation session, suggesting that this approach provides an effective treatment option for patients with unresectable large-volume liver lesions⁸⁷. In addition to liver tumours^{61,88–91}, MWA is also widely used for the ablation of tumours of the lungs⁹², bones⁹³ and pancreas⁹⁴.

High-intensity focused ultrasound

HIFU is the only truly non-invasive thermal therapy. Furthermore, no upper limit of tissue tolerance exists, enabling patients to receive repeat HIFU procedures as often as is necessary^{95,96}. During HIFU, ultrasound waves (typically 0.8–3.5 MHz) are focused on a specific focal zone using

a curved transducer, phased array or lens, leading to the highest wave pressure at a small target lesion with negligible off-target exposure. HIFU devices typically have a target area that is ellipsoidal in shape (1–3 mm in diameter and 8–15 mm in length); therefore, many of these target areas must be placed side by side to cover the entire tumour. HIFU has the advantage of avoiding the possibility of tumour seeding along the needle track during treatment, which can lead to the development of haematogenous metastases in patients receiving electromagnetic thermal ablation⁹⁷.

Two predominant principles, including temperature increment and cavitation, are highly relevant to the biological effects of HIFU ablation⁹⁸. Under the stresses created by ultrasound pressure, the vibration or rotation of molecules in the tissue could be intensified, which makes the collision of these molecules with others more frequent, resulting in some loss of energy owing to friction, also known as frictional loss. Moreover, heat can be produced by frictional loss following intramolecular collisions, which can cause an immediate (within seconds) increase in temperature and irreversible coagulative necrosis of target tissues. Cavitation refers to the vibration of bubbles in response to ultrasound pressure waves. These bubbles are generated

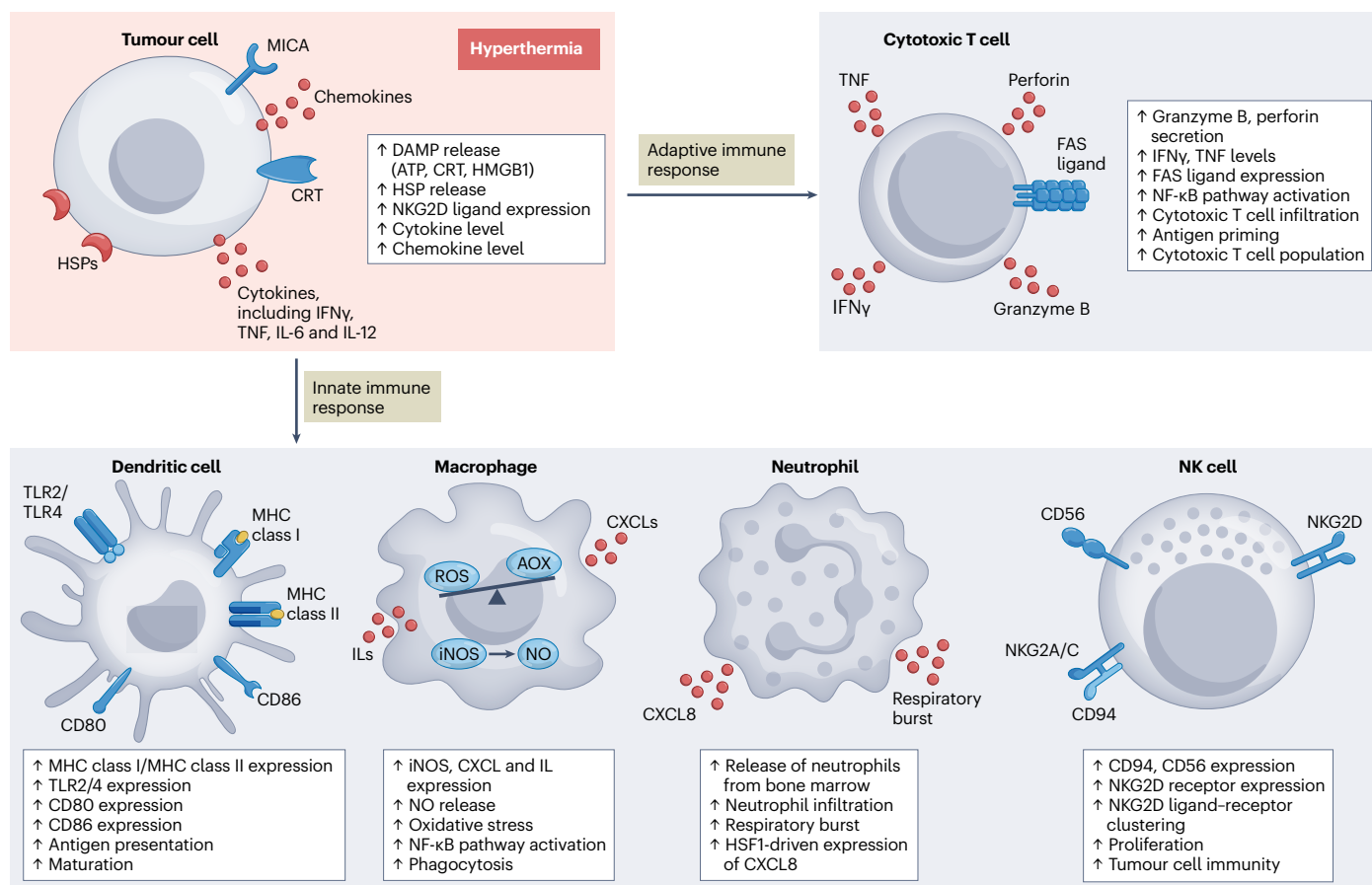


Fig. 1 | Effects of hyperthermia on innate and adaptive immunity.

Hyperthermia (typically 41–43 °C in most clinical applications) can induce immunogenic cell death of tumour cells, which results in the release of damage-associated molecular patterns (DAMPs, including ATP, calreticulin (CRT) and high mobility group box 1 (HMGB1)) and heat-shock proteins (HSPs), increases the expression of several biomarkers of immune cell activation, such as NKG2D and

MICA, and promotes the release of several pro-inflammatory cytokines and chemokines, which are able to further enhance the activation of different immune cells to boost both innate and adaptive immunity and promote tumour microenvironment remodelling. AOX, alternative oxidase; ILs, interleukins; iNOS, inducible nitric oxide synthase; NK, natural killer; NO, nitric oxide; ROS, reactive oxygen species; TLR, Toll-like receptor.

in a liquid when the tensile strength of the liquid is exceeded owing to the external ultrasound pressure waves. Where such waves reach sufficient frequency and amplitude, the liquid separates or fractures to form voids, which are then filled with air and/or the liquid itself. These voids, also known as gas nuclei, are rarified cavities that are able to further grow and collapse in response to more ultrasound waves, undergoing so-called stable cavitation and inertial cavitation. The stable cavitation caused by the oscillation of bubbles usually places mechanical stress on both vessel walls and cell membranes, resulting in improved extravasation of drugs into target tissues as well as their intracellular delivery. Inertial cavitation, generated by the collapse of bubbles upon ultrasound stimulation beyond a certain energy threshold, can introduce dramatic localized increases in temperature and shockwaves, leading to necrosis owing to mechanical stress and thermal invasion.

So far, the FDA has approved several HIFU devices, including the Sonablate and Ablatherm systems (transrectal HIFU) and the TULSA-PRO system (transurethral HIFU) for the ablation of primary prostate tumours^{99–101}, and the Sonalleve and ExAblate systems for osteoid osteomas and uterine fibroids, respectively^{30,102–104} (Table 2). HIFU can also be used for other indications, including breast cancers¹⁰⁵, desmoid tumours¹⁰⁶, liver cancers¹⁰⁷, pancreatic cancers¹⁰⁸ and kidney cancers¹⁰⁹. Device positioning for target tumours is usually guided by MRI, which can also provide the necessary real-time temperature feedback for robotically driven HIFU ablation^{110,111}. In an early clinical study, 70 patients with T1c–T2a cancers of any Gleason score limited to one side of the prostate received focal HIFU using the Sonablate system; 7 patients also received neoadjuvant androgen-deprivation therapy. Residual tumour material was detected in 11.9% and 18.4% of patients at 6 months and 12 months post-treatment, respectively. Adverse effects included urethral strictures and urinary tract infections in 8.6–4% and 11.4–4% of patients, respectively, depending on the area of the prostate that was ablated¹¹². Immediate tissue responses during HIFU can be observed using advanced technologies to evaluate therapeutic effectiveness. For example, tissue change monitoring is a quantitative software module that can be applied alongside the Sonablate system to evaluate the adequacy of energy delivery to each HIFU ablation site. In this system, a radiofrequency signal is sent to a targeted ablation site before and after HIFU with quantification based on comparisons of radiofrequency pulse-echo ultrasound signals at each HIFU lesion. Finally, an easy-to-read on-screen colour overlay with focal point markers is displayed to guide HIFU delivery. For the Sonalleve MR-HIFU system, which was approved for patients with osteoid osteoma of the extremities by the FDA in 2020, a self-contained cooling system is included in which cooled water is circulated within the ultrasound window during treatment, thus protecting tissues immediately surrounding the ablation zone.

Magnetic hyperthermia

Magnetic hyperthermia is another important thermal therapeutic modality that is currently in clinical use. In this method, a series of magnetic particles are prepared as transducers enabling the highly efficient conversion of electromagnetic energy into heat when subjected to an alternating magnetic field¹¹³. Nanoparticle-mediated hyperthermia is induced by the coupling of magnetic moments of the atoms in the nanoparticles within the imposed time-dependent external magnetic field (with a typical radiofrequency range of 100–300 kHz)^{114–117}. Nanoparticle-mediated magnetic hyperthermia can enhance the local temperature of the tumour within the range of 42–46 °C, resulting in cell death via either apoptosis or necrosis^{116,118}. Magnetic nanoparticles

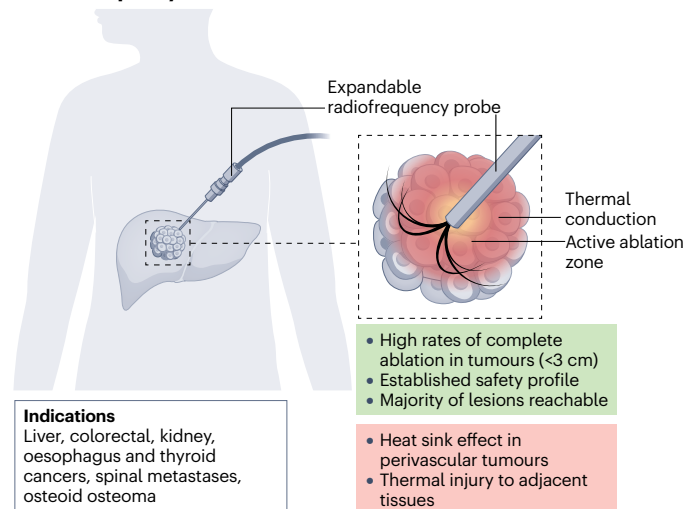
are administered intratumourally with the purpose of maximizing the intratumoural concentration of the nanoparticles, which is a crucial determinant of heat generation. In 2012, a magnetic hyperthermia device was approved by the EMA as an adjunct therapy for patients with recurrent glioblastoma who are also receiving radiotherapy¹¹⁹. In 2021, a clinical trial testing a similar approach for tumour ablation in men with prostate cancer received ethics approval (NCT05010759).

Combinations with immunotherapy

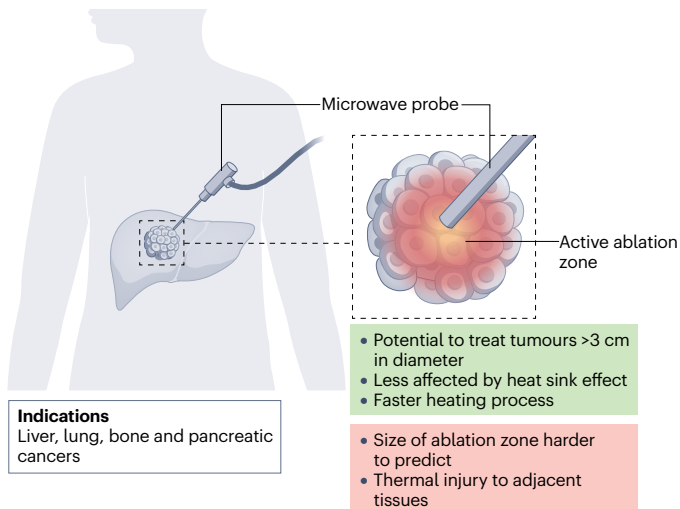
In addition to causing direct damage to tumour tissues, thermal therapies might also induce a systemic anticancer immune response, providing an opportunity to eradicate both primary tumours and their metastases (Fig. 1). The targeted thermal stress generated by such therapies can cause ICD, leading to the release of damage-associated molecular patterns such as calreticulin, heat-shock proteins (HSPs), ATP and high mobility group box 1 (HMGB1)^{18,120}. These damage-associated molecular patterns act as ‘find me’ signals (ATP and HMGB1) or ‘eat me’ signals (HSPs and calreticulin) that, with the help of chemokines, mobilize antigen-presenting cells and their precursors to tumours, resulting in the reversal of immunosuppression and establishing a favourable immunogenic TME^{121–124}. Thermal stimulation has also been shown to promote the differentiation of regulatory T (T_{reg}) cells into T helper 17 (T_H17) cells via hyperthermia-induced IL-6 secretion, which promotes antitumour activity in preclinical models¹²⁵. Similarly, CD8⁺ T cell function can be promoted by mild thermal stimulation as demonstrated by increased antigen-dependent activity and activation-induced IFN γ release at 39.5 °C relative to 33 °C and 37 °C, respectively, in preclinical models¹²⁶. Meanwhile, activation of other innate immune cells, including natural killer (NK) cells, macrophages and neutrophils, as well as dendritic cell (DC) maturation are promoted upon heating^{127–129}. For example, the cytotoxicity of NK cells can be substantially increased under thermal stress, which might be explained by HSP70 facilitating the upregulated expression of NKG2D, leading to improved recognition and destruction of tumour cells by NK cells^{130,131}. Hyperthermia also promotes the differentiation of CD8⁺ and CD4⁺ T cells into memory T cells and T_H cells, respectively, leading to long-term tumour-specific immune responses on tumour cell rechallenge in mouse models^{132,133}.

Nowadays, the main immunotherapies used in clinical cancer therapy are ICI and cellular immunotherapies such as CAR T cells. The therapeutic indices of these modalities are acceptable, although improvements in the performance of these therapies are important to expand the scope of their application. Heat stimulation alone can facilitate antitumour immunity¹³⁴; therefore, thermal therapies hold great promise to synergize with these clinically approved immunotherapies. Indeed, several clinical trials (either completed or ongoing) have investigated or are exploring the safety and efficacy of a thermal therapy combined with one or more immunotherapies, and some of these studies have already provided promising results (Table 3). For example, the anti-CTLA4 antibody tremelimumab combined with RFA resulted in an objective response rate (ORR) of 26% in 19 patients with advanced-stage unresectable HCCs with disease progression on sorafenib⁴¹. Progression-free survival at 6 and 12 months was 57.1% and 33.1%, and median time to progression and overall survival (OS) durations were 7.4 months and 12.3 months, respectively. Several patients had shrinkage of non-target lesions following a single round of intrahepatic nidus ablation and administration of tremelimumab, suggesting an immune-sensitizing effect of RFA. Moreover, no clear differences in the incidences of adverse events emerged across patients

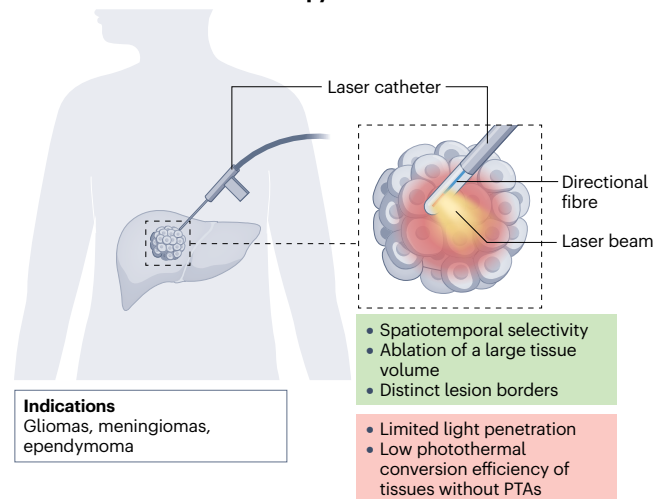
a Radiofrequency ablation



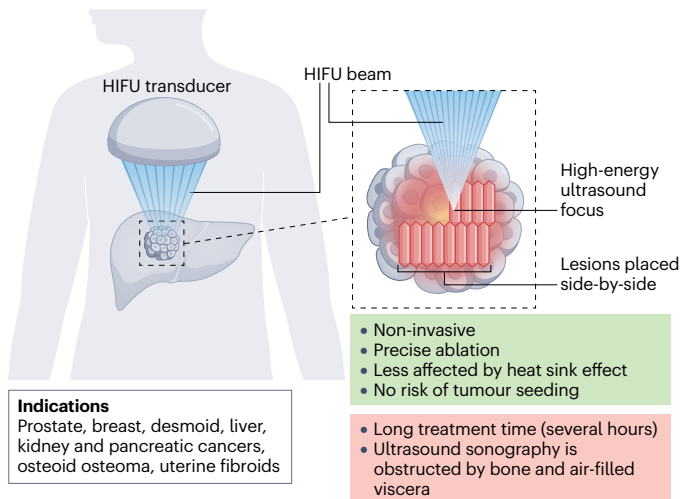
b Microwave ablation



c Laser interstitial thermal therapy



d HIFU ablation



e Magnetic hyperthermia

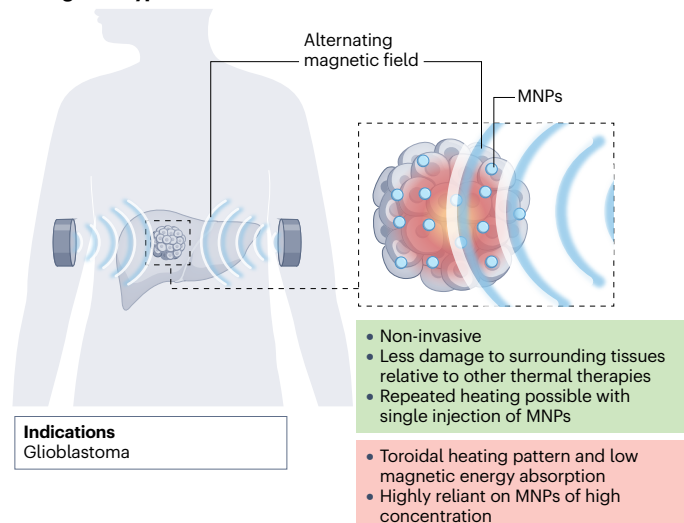


Fig. 2 | Graphical representation of the main thermal therapy modalities.

Several thermal therapies are available and are applied in certain clinical scenarios, each with its own specific characteristics. These include radiofrequency ablation

(part **a**), microwave ablation (part **b**), laser interstitial thermal therapy (part **c**), magnetic hyperthermia (part **d**) and high-intensity focused ultrasound (HIFU) ablation (part **e**). MNPs, magnetic nanoparticles; PTAs, photothermal agents.

receiving different doses of tremelimumab. The most common grade 3–4 adverse event was increased systemic aspartate aminotransferase (in 21% of patients)⁴¹. In another ongoing phase I/II trial involving a similar cohort of 48 patients with advanced-stage HCC, patients received either monotherapy with the anti-PD-1 antibody toripalimab or thermal ablation followed by toripalimab either 3 or 14 days post-ablation. At the latest follow-up cut-off, patients undergoing thermal ablation had ORRs of 37.5% and 31.2%, respectively, which were higher than that of patients receiving toripalimab monotherapy (18.8%). Safety results were also reported, indicating that 75% of patients had at least one any-grade adverse event. Grade 3–4 adverse events occurred in 18.7% of patients³⁹. Similarly, in 33 patients with advanced-stage HCC receiving anti-PD-1 antibodies (pembrolizumab or nivolumab) followed by RFA or MWA, the ORR increased from 10% to 24% following thermal ablation; however, whether this finding reflects synergy or a delayed response to immune-checkpoint inhibition remains uncertain. Median progression-free survival and OS durations were 5 months and 16.9 months, respectively⁴⁰. In summary, the safety profile of a thermal therapy–immunotherapy combination appears similar to that of single-agent ICIs. Further investigations of the efficacy of thermal therapy–immunotherapy combinations in patients with HCC should involve comparisons with the current standard-of-care approach.

Thermal immuno-nanomedicines

On the basis of data described in the previous section, thermal therapy–immunotherapy combinations have the potential to improve outcomes^{41,134}. However, ICIs have several limitations, including the risk of immune-related adverse effects and the lack of responses in most patients, both of which remain major challenges for such combinations.

For example, in a phase III clinical trial completed in 2019, 609 patients with stage III NSCLC received the anti-PD-L1 antibody atezolizumab; 64% of patients had treatment-related adverse events of any grade and 32% had clinically serious events¹³⁵. Only 18% of patients had an objective response.

With the development of nanotechnology, nanomedicine arises as a promising way to address these limitations owing to several advantages over more traditional therapies, including increased intratumour accumulation, prolonged in vivo circulation time, and the potential for improved safety and/or controlled release of their pharmacological cargoes. At least 20 nanomedicines are currently in clinical use as cancer therapies globally, and 566 clinical trials involving nanomedicines as cancer therapies were conducted between 2016 and 2020 (ref. 136). Some of these agents (such as nab-paclitaxel) had improved efficacy when combined with immunotherapy^{14,137,138}. Nanomedicines also have the potential to combine multiple functions in a single platform, including the spatiotemporal synchronized co-delivery of hyperthermia and an immune adjuvant^{47,48,139}. Thus, the application of nanomedicines for thermal immunotherapy holds great promise (Fig. 3).

In situ vaccines

Vaccines have an important role in modern public health. Several different types of vaccines have been developed for various infectious diseases (including live-attenuated vaccines, mRNA vaccines and recombinant vector vaccines), and some of these designs have been applied to the development of therapeutic vaccines for patients with cancer^{140–145}. Most available cancer vaccines are designed to promote an immune response to a limited number of antigens rather than to all potential antigens derived from the target tumour,

Table 3 | Data from trials testing thermal therapy–immunotherapy combinations

Trial	Patient characteristics	Intervention	Outcomes
RFA			
NCT01853618 (phase I/II) ⁴¹	32 patients with HCC not amenable to potentially curative liver transplantation, resection or ablation	Tremelimumab every 4 weeks, subtotal RFA or CA on day 36	ORR: 26.3% among 19 evaluable patients; 6-month and 12-month PFS: 57.1% and 33.1%, respectively; median OS: 12.3 months; grade 3–4 serum AST increases in 21.8% of patients
NCT03695835 (observational) ²⁷⁸	27 patients with metastatic solid tumours, including 21 with prostate cancers	In situ cryosurgical lysis of tumour cells followed by intratumoural injections of pembrolizumab, nivolumab or ipilimumab plus sargramostim	ORR: 47% among 19 evaluable patients with prostate cancer; 62% of patients had post-therapy serum PSA reductions of >50%; grade 3–4 adverse events in 19% of patients
NCT03939975 (phase II) ⁴⁰	50 patients with advanced-stage HCC with disease progression on sorafenib or who were unable to tolerate sorafenib monotherapy	At least one cycle of pembrolizumab or nivolumab monotherapy; 33 patients (66%) received subtotal RFA repeated up to 4 times	ORR: 24%; median PFS: 5 months; median OS: 16.9 months; clinically serious adverse events in 14% of patients
RFA/MWA			
NCT03864211 (phase I/II) ³⁹	48 patients with advanced-stage HCC with disease progression on at least one systemic therapy	Toripalimab monotherapy (arm A), subtotal ablation of up to 5 lesions plus toripalimab 3 days after ablation (arm B) or toripalimab 14 days after ablation (arm C)	ORR: 18.8% in arm A, 37.5% in arm B and 31.2% in arm C; grade 3–4 TRAEs in 18.7% of patients in arm A and 25.0% in arms B and C, respectively.

NCT04707547, NCT03101475, NCT02851784 are listed as ‘completed’ in ClinicalTrials.gov, with results unavailable, to the best of the authors’ knowledge. AST, aspartate aminotransferase; CA, chemotherapy ablation; HCC, hepatocellular carcinoma; MWA, microwave ablation; ORR, objective response rate; OS, overall survival; PFS, progression-free survival; PSA, prostate-specific antigen; RFA, radiofrequency ablation; TRAEs, treatment-related adverse events. *Currently listed as ‘ongoing’ in ClinicalTrials.gov.

which might explain their limited effectiveness. As described previously, the ICD induced by tumour-ablative thermal therapies causes the release of various tumour-associated antigens (TAAs), which could, if recognized by the immune system, lead to an antitumour immune response capable of suppressing the growth of both primary tumours and their metastases¹⁸. Thus, TINs could act as an in situ cancer vaccine.

To test this hypothesis, investigators synthesized an amphiphilic polymer, glycol chitosan-graft-polyaniline, and used it to encapsulate the Toll-like receptor 7/8 agonist resiquimod as a nanoparticle-based in situ cancer vaccine¹⁴⁶. In an acidic TME, this nanoparticle system induced a localized increase in temperature following near-infrared (NIR) laser irradiation. This mild hyperthermia promoted the expression of HSP70 and activation of the immune system, which

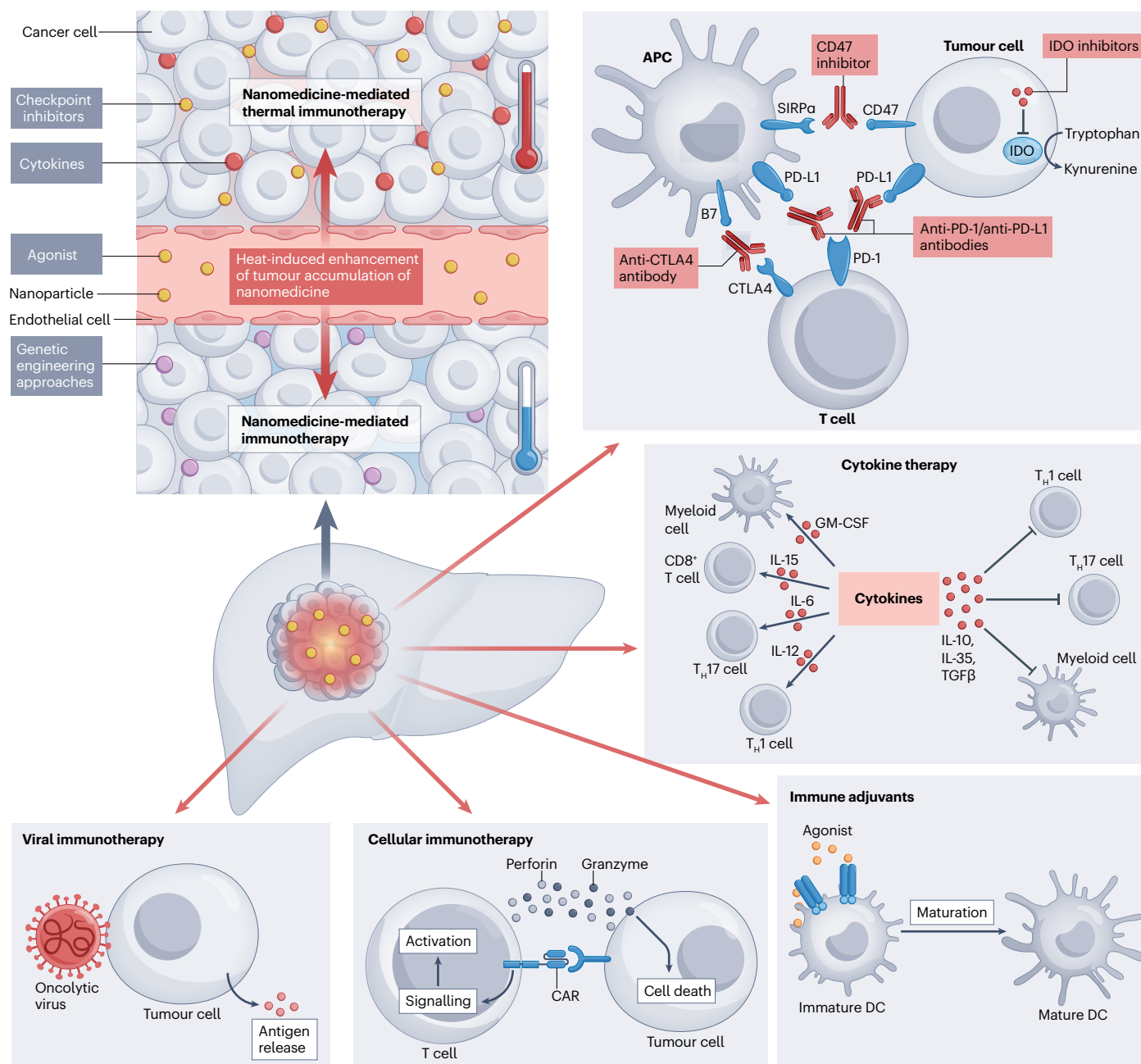


Fig. 3 | Thermal immuno-nanomedicines. Nanomedicines for thermal immunotherapy are designed to induce targeted tumour-specific hyperthermia and to synergize with the various clinically approved immunotherapies (including vaccines, immune-checkpoint inhibitors, cellular immunotherapies, oncolytic viruses and cytokine therapies) to further improve their therapeutic

efficacy and potentially expand the scope of their application. APC, antigen-presenting cell; CAR, chimeric antigen receptor; DC, dendritic cell; GM-CSF, granulocyte-macrophage colony-stimulating factor; IDO, indoleamine-2,3 dioxygenase; SIRP α , signal-regulatory protein- α ; T_H, T helper.

synergized with resiquimod to further strengthen antitumour immunity in mouse models with pre-established subcutaneous tumours. Exposure to the TIN resulted in complete tumour regression in 43% of mice; in comparison, monotherapy with either mild hyperthermia or resiquimod delayed tumour growth but failed to induce tumour regression. Cancer cell membrane-coated nanomedicines have attracted considerable research interest as cancer vaccines owing to their selective intratumour accumulation due to homotypic aggregation and the potential to present multiple antigens, thus overcoming the issue of tumour heterogeneity^{147–149}. In a hybrid design, bacterial outer membrane vesicles were mixed with cancer cell membranes to formulate a membrane-camouflaged poly(lactic-co-glycolic) acid–indocyanine green (PLGA-ICG) nanoplateform¹⁵⁰. This nanoplateform features several natural adjuvant components inherited from the parent bacteria as well as cell-surface TAAs from the cancer cells. More importantly, the localized increase in temperature induced by ICG under NIR irradiation could generate supplementary TAAs to further stimulate the immune system. When tested in mouse models of subcutaneous melanoma, this TIN was found to confer an 80% reduction in tumour volume, which is substantially greater than that achieved with monovesicle-camouflaged nanoparticles (30%), indicating that simultaneous PTT substantially improves the antitumour activity of immunotherapy. Future TIN-based cancer vaccines could include additional agents designed to alleviate the immunosuppressive effects of the TME. Elsewhere, investigators prepared a hydrogel loaded with ICG (a photothermal agent) and JQ1 (a BRD4 inhibitor) and tested this approach as post-surgical immunotherapy in mouse xenograft models¹⁵¹. This cancer vaccine can be activated and triggered by NIR irradiation for the on-demand release of tumour-specific antigens and JQ1, which can suppress disease relapse by facilitating DC maturation, enhancing cytotoxic T cell infiltration into tumours and inhibiting PD-L1-dependent immune evasion. TIN-based cancer vaccines integrate the ability to generate local hyperthermia and to modulate the immune system for improved antitumour activity and provide a versatile platform for the rational design of more sophisticated therapeutic systems. The combination of hyperthermia and one or more immune adjuvants promotes immune cell infiltration and confers durable antitumour immunity in mouse models, suggesting the release of multiple TAAs from the primary tumour^{150,151}. Use of this *in situ* vaccine approach provides a powerful and personalized method that avoids the need to identify and manufacture TAAs. Although promising, this technology is still currently only being tested in preclinical investigations.

Immune-checkpoint inhibition

Immune checkpoints are important cell-surface immunoregulatory signalling proteins that can be hijacked to enable cancer cells to evade the immune system^{152,153}. Thus, immune-checkpoint inhibition, enabling T cells to recognize and kill cancer cells, is now an important therapeutic modality. Currently, several ICIs, such as those targeting PD-1, PD-L1, CTLA4 or CD47, are used clinically to treat various forms of cancer. Furthermore, agents targeting enzymes with immunosuppressive effects, such as indoleamine-2,3-dioxygenase 1 (IDO1), are an area of considerable research interest^{154–157}. Preclinical evidence suggests that combination with hyperthermia further enhances the anticancer activity of these agents.

CTLA4. CTLA4 expressed on effector T (T_{eff}) cells and T_{reg} cells competes with CD28 for B7 ligands and, when activated, suppresses the activation and growth of T_{eff} cells and the generation of effector memory

T (T_{em}) cells^{158,159}. Thus, inhibition of CTLA4 signalling can restore the activity of T_{eff} cells and facilitate the production of T_{em} cells, promoting antitumour immunity¹⁶⁰. Data from an increasing number of studies indicate that the anticancer activity of anti-CTLA4 antibodies can be improved by combination with hyperthermia^{161–164}.

Anti-CTLA4 antibodies have been combined with magnetic hyperthermia and PTT in several preclinical studies. In one example, investigators prepared bullet-shaped Fe_3O_4 -tipped mesoporous silica magnetic nanoparticles (also known as Janus nanobullets) with a chlorine e6 (Ce6) cytotoxic payload enveloped by a cancer cell-derived membrane. This Janus nanobullet enabled TME-responsive Ce6 release in the presence of an oscillating magnetic field and NIR irradiation, leading to glutathione depletion, reactive oxygen species (ROS) augmentation and magnetic hyperthermia, with evidence suggestive of ICD in mouse xenograft models of breast cancer¹⁶¹. Elsewhere, researchers have leveraged the excellent photothermal properties of carbon nanotubes, especially single-walled nanotubes, for targeted thermal therapy. Single-walled nanotubes non-covalently modified with polyethylene glycol (PEG) were constructed and used for PTT in combination with anti-CTLA4 antibodies, resulting in a significantly longer time to cancer recurrence in mouse models of lung metastases. This approach facilitated T_{eff} cell expansion while also substantially reducing the number of T_{reg} cells in distant lesions¹⁰⁵.

PD-1–PD-L1. The PD-1–PD-L1 signalling pathway is another important immune checkpoint. PD-1 is predominantly expressed on activated T cells and its ligands, PD-L1 and PD-L2, are expressed on antigen-presenting cells and some cancer cells, which might explain the adaptive immune resistance, or immune evasion, seen in many cancers¹⁶⁵. Worldwide, at least ten different anti-PD-1 or anti-PD-L1 antibodies are in clinical use for the treatment of a wide range of cancers, including NSCLC and melanoma^{166,167}. As discussed previously, local hyperthermia promotes immune cell infiltration and induces ICD; thus, similar to anti-CTLA4 antibodies, thermal therapies are also likely to synergize with anti-PD-1 and anti-PD-L1 antibodies. To test this hypothesis, investigators prepared an injectable lipid gel loaded with PTAs and anti-PD-L1 antibodies. This temperature-sensitive lipid gel underwent phase transition and released the anti-PD-L1 antibody simultaneously with hyperthermia, which resulted in impaired tumour growth and T cell recruitment to the tumour⁴⁸. Similarly, when combined with anti-PD-L1 antibodies, ferrimagnetic vortex-domain iron oxide nanorings capable of magnetic hyperthermia impaired the growth of non-target lesions following thermal ablation of a target lesion in a mouse xenograft model. This approach conferred marked increases in CD3 and CD45 levels (indicating increased T cell and haematopoietic cell counts) and a considerable reduction in myeloid-derived suppressor cell levels in tumours distant to the target lesion, suggesting activation of systemic immunity¹⁶⁸.

As discussed above, combining anti-PD-1/anti-PD-L1 antibodies with hyperthermia is the most widely used immunotherapy–thermal therapy combination strategy^{169,170}. However, as mentioned previously, the systemic administration of these antibodies as monotherapies typically results in limited response rates and risk of immune-related adverse effects¹⁷¹. To address this problem, gene-editing or gene-silencing technologies could be utilized to achieve persistent inhibition of PD-1–PD-L1 signalling. Small interfering RNAs (siRNAs) theoretically enable the expression of any specific gene to be silenced by post-transcriptional mRNA degradation, providing a powerful technology that could potentially be adapted for cancer therapy^{172,173}. However, clinical approaches involving siRNA are often impaired by rapid

enzymatic degradation and an inability to penetrate cell membranes; therefore, most siRNA-based therapies require a targeted delivery system. An example is provided by patisiran, an siRNA-containing lipid nanoparticle approved by the FDA in 2018 for the treatment of peripheral nerve disease caused by hereditary transthyretin-mediated amyloidosis in adults. Delivery of the siRNA using a lipid nanoparticle improved the in vivo stability and enabled selective accumulation in the liver owing to surface adsorption of apolipoprotein E (ApoE)¹⁷⁴. Other promising siRNA-containing nanoparticles currently in development include siRNA-GalNAc conjugates and Dynamic PolyConjugates, which involve the conjugation of siRNAs with specific ligands or polymers, respectively¹⁷⁵. Among other possibilities, TINs could provide the delivery vehicle for PD-L1-silencing siRNAs. In a preclinical study, gold nanoparticles capable of both delivery of PD-L1 siRNA and PTT were prepared and shown to suppress the proliferation of cancer cells in mouse xenograft models through PD-L1 knockdown and hyperthermia following NIR laser irradiation¹⁷⁶. The CRISPR–Cas9 system provides an alternative target-specific gene-editing tool^{177,178}. Since its discovery in 2010, CRISPR–Cas9 has been widely used for in vitro gene editing in experimental models, and this approach theoretically has tremendous potential as a treatment of various diseases owing to its versatility and simplicity^{179–182}. As a powerful gene-editing tool, this technique creates considerable ethical and safety concerns, especially when used in applications such as in vivo germline gene editing; nonetheless, considerable public support and research interest exist in the therapeutic application of this technology. Early proof-of-principle data supporting the in vivo use of CRISPR–Cas9 technology in patients come from a phase I study in which six patients with hereditary transthyretin-mediated amyloidosis and polyneuropathy received a lipid nanoparticle formulation containing RNA encoding Cas9 and a guide RNA targeting *TTR*. Early results showed durable suppression of the target gene (mean reduction in TTR protein levels of 52%) after a single dose with only mild (grade 1) adverse events¹⁸³. CRISPR–Cas9 has been applied to the development of TINs designed to also disrupt *PD-L1*. In a preclinical study, investigators fabricated supramolecular cationic-gold nanorods loaded with plasmids encoding *Cas9* and a single guide RNA targeting *PD-L1* downstream of a heat-sensitive promoter, which potentially reduces the risk of off-target events compared with untargeted expression¹⁸⁴. Upon NIR II (1,000–1,700 nm) laser irradiation, the mild hyperthermia generated by PTT increased the local temperature to 42 °C, enabling transcriptional activation of *Cas9* and suppression of *PD-L1*, which promoted T cell infiltration and an expansion of T_{em} cells, thus providing systemic antitumour immunity that delayed the growth of distant lesions in mouse xenograft models.

CD47. CD47 is expressed on different types of cancer cell, such as non-Hodgkin lymphoma, acute myeloid leukaemia, glioblastoma, ovarian, breast, colon, bladder, hepatocellular, and prostate cancers, and binds to signal-regulatory protein- α (SIRP α) on macrophages to suppress phagocytic function, acting as a ‘do not eat me’ signal¹⁸⁵. Anti-CD47 antibodies have been shown to be well tolerated in a phase I clinical trial involving 62 patients with advanced-stage cancers¹⁸⁶. In another phase Ib study, the combination of an anti-CD47 antibody with rituximab showed promising activity in patients with relapsed and/or refractory non-Hodgkin lymphoma without the emergence of any clinically significant adverse events¹⁸⁷. Several more clinical trials involving anti-CD47 monoclonal antibodies as cancer treatments are currently ongoing (NCT02216409, NCT02367196, NCT02447354 and NCT02488811)^{136–138}. Data from several studies also demonstrate that

anti-CD47 antibodies facilitate DC activation and T cell priming¹⁸⁸. Similar to the experience with other ICIs, concurrent hyperthermia potentiates the antitumour activity of anti-CD47 antibodies in preclinical models^{189–192}. An example of this effect is provided by a hybrid therapeutic nanocarrier fusing CD47-overexpressing exosomes with ICG and imiquimod-loaded thermoresponsive liposomes¹⁹⁰. This approach promoted DC maturation and the lethality of CD8⁺ and CD4⁺ T cells and macrophages in a mouse xenograft model.

IDO1 inhibition

IDO1 catalyses the metabolism of tryptophan, which is the first and rate-limiting step in the kynurenine synthesis pathway^{193–196}. This enzyme enables cancer cells to evade the immune system through two main mechanisms: tryptophan depletion, which can suppress the activation and proliferation of T_{eff} cells, and kynurenine production, which is cytotoxic to T cells and NK cells but promotes the proliferation of T_{reg} cells and T_H17 cells. Thus far, more than 10 different IDO1 inhibitors have been tested in preclinical studies or clinical trials, albeit without any notable success in phase III studies¹⁹⁷. Several studies have focused on combining IDO1 inhibitors with thermal therapy in an attempt to improve the efficacy of these agents^{198,199}. Ultrathin microporous polymer nanosheets conjugated to a NIR-sensitive semiconducting polymer and coated with the IDO1 inhibitor 1-methyltryptophan (1-MT) were used to simultaneously induce ICD and inhibit IDO1 activity²⁰⁰. This system enables the concurrent generation of hyperthermia, ROS and oxygen, which alleviates tumour hypoxia and elicits the generation of ROS. After intravenous administration and NIR laser irradiation, mice injected with TINs had a substantial reduction in tumour growth, whereas mice injected with the nanosheet without 1-MT plus irradiation had moderate inhibition of tumour growth and non-irradiated mice had minimal inhibition of tumour growth. Our group synthesized a thermal-sensitive nitric oxide (NO) donor poly(acrylamide-co-acrylonitrile-co-vinylimidazole)-S-nitrosothiol pendant copolymer with an upper critical solution temperature (42.1 °C in a weakly acidic TME), which was used to prepare an erythrocyte membrane-camouflaged nanoparticle with the NIR II photothermal agent IR1061 and with 1-MT as a payload¹³⁹. The hyperthermia at the tumour site induced by PTT elicited ICD but also triggered the release of 1-MT and NO to regulate the kynurenine metabolic pathway and alleviate hypoxia, respectively^{201–203}. This multifunctional nanomedicine promoted the recruitment of cytotoxic T cells to the tumour site and suppressed PD-L1 expression and T_{reg} cell proliferation, while polarizing tumour-associated macrophages towards an M1 phenotype, collectively reversing immunosuppression in hypoxic and immune-cold tumours in a mouse xenograft model. In addition to synergy between hyperthermia and IDO1 inhibition, the controlled release of NO from TINs could remodel the TME and further enhance antitumour activity.

Cellular immunotherapy

The aim of cellular immunotherapy is to isolate immune cells, either from patients with cancer or from HLA-matched or potentially even unmatched individuals, genetically modify and/or expand the cells in vitro, and then infuse them back into the patient to directly kill the tumour and/or promote antitumour immunity²⁰⁴. Several different forms of cellular immunotherapies exist, including tumour-infiltrating lymphocytes, TCR-engineered T cell and CAR T cell therapies, and genetically modified and/or expanded NK cell or DC-based immunotherapies^{205–207}. Currently, five different CAR T cell constructs are approved for patients with B cell malignancies (idecabtagene vicleucel, lisocabtagene maraleucel, tisagenlecleucel, brexucabtagene autoleucel

and axicabtagene ciloleucel) and two for multiple myeloma (idecabtagene vicleucel and ciltacabtagene autoleucel)²⁰⁸. Anti-CLL1 CART cells are also approved by the FDA as orphan drugs for patients with acute myeloid leukaemias²⁰⁹. Various other constructs are also currently in the late stages of clinical testing, including relmacabtagene autoleucel as a second-line therapy for patients with diffuse large B cell lymphoma²¹⁰. Cellular immunotherapies are clearly effective in patients with advanced-stage B cell malignancies; however, limited infiltration of the re-infused cells is often seen in patients with solid tumours receiving experimental CAR T cells and this restricts the broader use of cellular immunotherapies. Ablative hyperthermia has the potential to damage or destroy extracellular matrix and other TME components, thus removing a major barrier to infiltration^{211–214}. The ensuing ICD could also stimulate the production of pro-inflammatory cytokines and chemokines, and promote the recruitment of both adoptive immune cells and potentially native immune cells to non-ablated lesions^{214,215}. For example, adoptive transfer of gp100-specific pmel T cells following hollow gold nanoshell-mediated PTT enables antitumour activity against non-ablated lesions as demonstrated by the suppression of lung metastases in a mouse xenograft model of melanoma²¹⁶. A similar approach was investigated using PTT to improve the antitumour activity of CAR T cells in a mouse xenograft model of melanoma²¹⁴. In this study, ICG-loaded PLGA nanoparticles were used to induce local hyperthermia following laser irradiation leading to ICD of cancer cells, resulting in the destruction of the extracellular matrix, immune cell infiltration and increased tumour-specific expression of several chemokines and cytokines. T cells genetically engineered to express a CAR targeting chondroitin sulfate proteoglycan 4 (CSPG4), a specific antigen overexpressed in glioblastoma and melanoma, were infused 2 h after laser irradiation. The tumour-specific accumulation of anti-CSPG4 CAR T cells was increased in mice previously exposed to PTT, resulting in substantial suppression of tumour growth up to 20 days following CAR T cell administration. These preclinical data suggest that combining mild hyperthermia with CAR T cell infusion enhances the antitumour activity of CAR T cells. Mild hyperthermia induced by NIR light not only inhibits the growth of cancer cells but can also enable the controlled *in vivo* generation and/or release of therapeutics. To further assess this possibility, investigators constructed and screened panels of synthetic thermal gene switches containing the combinations of heat-shock elements and core promoters to identify constructs that respond to mild hyperthermia²¹⁷. Under photothermia generated using PTT (40–42 °C), these heat-responsive promoters are able to control the production of several immunostimulatory genes, including those encoding bispecific T cell engagers and IL-15 superagonists, with antitumour activity observed in mouse models of antigen-negative tumours. Elsewhere, biocompatible electrospun nanofibres embedded with light-sensitive iron oxide nanoparticles were used to promote the photo-activated *in vitro* permeabilization of T cells to CRISPR–Cas9 ribonucleoprotein complexes and siRNAs targeting PD-1 (ref. 218). This strategy did not affect the proliferation or phenotype of the engineered T cells, with antitumour activity similar to that observed with PD-1-expressing CAR T cells plus anti-PD-1 antibodies.

Cytokine-based therapies

Cytokines are a category of signalling molecules that includes interferons, chemokines, lymphokines and interleukins with an essential role in regulating the proliferation and activation of immune cells²¹⁹. Approximately 200 different cytokines have been identified as having roles in cell signalling and several, such as IL-7 and IL-12, are currently being tested in

clinical and preclinical studies^{220,221}. However, systemic administration of cytokines as monotherapies is usually associated with dose-limiting toxicities, which hinders the use of unmodified cytokines as cancer therapies²²². Among other approaches, researchers have attempted to use nanotechnology to improve the safety, bioavailability and therapeutic efficacy of systemically administered cytokines. An early example of a nanomedicine designed for targeted delivery of a cytokine payload is provided by CYT-6091, a PEGylated colloidal gold nanoparticle with TNF conjugated to its surface. This method of delivery avoids hypotension, which is a dose-limiting toxicity associated with unmodified TNF²²³. Two phase I clinical trials testing the safety and tolerability of CYT-6091 have been completed (NCT00356980 and NCT00436410) and a clinical trial agreement on a phase II study has been signed between the manufacturer and the US National Cancer Institute^{224,225}. The rationale for combining these two modalities is based on the ability of hyperthermia to elicit ICD and promote transfection with cytokine-encoding plasmids, leading to the localized release of cytokines capable of TME remodelling. In one approach, protamine-modified superparamagnetic iron oxide nanoparticles capable of magnetic hyperthermia were combined with TNF-encoding plasmids²²⁶. This nanoparticle demonstrated robust transfection efficiency, possibly assisted by the elevation of temperature created by the alternating magnetic field, with promising synergistic antitumour activity in a mouse xenograft model of HCC. In another approach, CuS–SiO₂ nanoparticles were conjugated with positively charged poly((2-dimethylamino)ethyl methacrylate), forming complexes with negatively-charged IL-12-encoding plasmid DNA²²⁷. Under NIR II irradiation, hyperthermia induced by CuS–SiO₂ significantly improved the efficiency of transfection, leading to improved IL-12 release and antitumour activity beyond the target lesion. In the bilateral B16F10 mouse xenograft model used, this TIN inhibited the growth of tumours located close to the injection site and suppressed distant lesions within the same mouse, suggesting the emergence of an abscopal effect.

Oncolytic viruses

Oncolytic viruses are a novel immunotherapy modality involving the use of either naturally occurring or genetically modified viruses to selectively infect and eliminate cancer cells via ICD^{228,229}. Thus far, three oncolytic viruses have been approved for clinical use globally: H101 for head and neck cancer in China; talimogene laherparepvec for melanoma in the USA and Europe; and tesoratuvir for malignant glioma in Japan^{230,231}. At least 10 other oncolytic viruses are currently being tested in clinical trials. In an early attempt to explore the antitumour activity of oncolytic viruses in combination, investigators combined viral oncolysates obtained from individual patients with autologous DCs with modulated electrohyperthermia, which involves selective heating of the extracellular matrix of malignant tissues²³². In this approach, the immune system of patients is pre-conditioned via injection of an avian Newcastle disease virus and hyperthermia using a radiofrequency of 13 MHz. This step is followed by the administration of autologous DCs exposed to viral lysates obtained by exposing the patient's tumour cells to the Newcastle disease virus *in vitro*. Preliminary investigations indicate that this approach confers a median OS duration of 30 months in a retrospective cohort of 10 patients with newly diagnosed glioblastoma.

Future prospects and challenges

On the basis of the available preclinical data, TINs combining thermal therapies and immunotherapies have the potential to improve therapeutic efficacy. The data suggest two main mechanisms of action of

TINs: relieving the immunosuppressive effects of the TME and promoting antitumour immunity via ICD. Thus far, approaches combining these two modalities in a single nanoparticle have not been investigated in clinical trials and several challenges continue to hinder clinical translation.

Firstly, many thermal therapies, including TINs, are affected by a common problem, known as the ‘heat-sink effect’ or convective cooling, which refers to heat being carried away from the target via the circulation, thus compromising attempts to increase only the local temperature⁸³. For RFA in particular, this effect creates an active heating zone that is limited to several millimetres surrounding the tip of the applicator. Heat conduction has an important role in expanding the heating area, which explains the greater susceptibility of this technique to the heat-sink effect compared with other thermal therapeutic modalities²³³. Moreover, the need for homogenous hyperthermia across the entire target lesion creates a further clinical challenge. The deposition of energy during interstitial heating is limited to the close proximity of the probes owing to the line/point nature of interstitial heat sources. As a result, multiple small applicators must be arranged and spaced 1–2 cm apart in order to offset heat loss and cover the entire target lesion, which complicates both clinical operation and ablation zone prediction. Therefore, a trend towards the design of more advanced applicators that provide greater spatial control of energy deposition to ensure ideal temperature distributions across the target tumours while sparing the neighbouring non-malignant tissues is currently emerging. Regardless of these limitations, most TINs currently in preclinical development are activated by lasers for various reasons. Photothermal activation is an appealing method that enables the precise, targeted activation of TINs. Furthermore, lasers are more amenable to laboratory experiments involving small animals relative to other possible methods of TIN activation. However, the limited tissue penetration depth associated with optical activation (restricted to within a few centimetres of the skin surface) impairs the widespread implementation of this approach. Endoscopes equipped with optical fibres designed to deliver light to deep tissues have the potential to address this issue, which would considerably improve the potential for clinical implementation. In this regard, low-temperature-sensitive, liposome-encapsulated nanobubbles have already been demonstrated to enhance the performance of HIFU ablation^{234,235}, and this approach has the potential to enable localized activation of TINs.

Precise control of temperature is another major challenge associated with the use of TINs. The ICD induced by hyperthermia is highly dependent on achieving an optimal temperature, which means the ‘more is better’ paradigm might be inappropriate for hyperthermia designed to promote the efficacy of immunotherapy²³⁶. Furthermore, the higher temperatures required for the thermal ablation of tumour cells might damage non-malignant tissues, including immune cells. This approach can also restrict intratumoural blood flow and cause the tumour to form a barrier that prevents immune cell infiltration and immune induction²³⁷. However, owing to the disordered geometry and seemingly random location of the tumour blood vessels, uniform heating of an entire tumour is often difficult to achieve. Although various thermometry techniques based on different imaging modalities (such as MRI or photoacoustic imaging) have been applied clinically, each method still has certain limitations^{238,239}. For example, hard-wire thermistor or thermocouple-based sensors are prone to measurement errors arising from electromagnetic interference, which might introduce artefacts on MRI images, leading to inaccurate temperature monitoring. Photoacoustic imaging can also provide relative

temperature measurements; however, obtaining accurate measurements of absolute temperature without understanding the baseline temperature is challenging, particularly in deep tissues with unknown optical and acoustic properties. Owing to strong light attenuation in biological tissues, photoacoustic imaging can only achieve sufficient contrast from endogenous chromophores at tissue depths of <4 cm (ref. 240). Temperature monitoring is essential to precisely control the thermal conditions of the tumour and enable optimal treatment efficacy with minimal adverse effects; thus, an urgent need exists to improve the technology used for heating and temperature monitoring.

A number of different immunotherapies are currently available. Therefore, how best to select the most appropriate immunotherapy for use in combination with a specific thermal therapy and/or incorporation into TINs in order to maximize therapeutic efficacy remains a major challenge. The rapid development of artificial intelligence (AI) technology provides one possible solution to this problem. In addition to the development of new treatment methods and drugs, the massive amount of medical data available can be analysed using big data analysis and AI technology to achieve the goal of personalized medicine^{241–243}. For example, IBM, in collaboration with Memorial Sloan Kettering Cancer Center, designed an AI assistant decision-making system, named Watson for Oncology (WFO)²⁴⁴. This system was trained for >4 years based on the national comprehensive cancer network (NCCN) guidelines and >100 years of treatment experience and is designed to suggest the optimal therapeutic schedule for each individual patient. Thus, once a larger body of clinical data on the performance of thermal therapy–immunotherapy combinations, including TINs, becomes available, such approaches could be used to analyse these data and identify predictors of therapeutic efficacy that enable the most appropriate thermal immunotherapy protocol to be recommended on an individual basis.

Meanwhile, despite the impressive outcomes that have been achieved with TINs in preclinical studies, the clinical application of such nanomedicines remains in the proof-of-concept phase and has some limitations in clinical translation. Firstly, the enhanced permeability and retention (EPR) effect has long been believed to be an important mechanism for the accumulation of nanomedicines in tumour tissues. However, the significance and even the existence of the EPR effect in solid tumours in patients is becoming a controversial topic²⁴⁵. Emerging evidence suggests that nanoparticles are able to enter solid tumours through mechanisms more complex than previously thought, potentially going beyond simple extravasation through gaps in the endothelial lining. For example, a recent report demonstrated that the EPR effect might not be the major underlying mechanism of nanoparticle extravasation and that active transcytosis also contributes to the tumour-specific accumulation of nanoparticles^{245,246}. Therefore, innovative strategies designed to enhance the tumour-specific accumulation of nanoparticles will be needed to facilitate the clinical translation of nanomedicines. Possible strategies include the use of physical methods such as hyperthermia, radiotherapy and ultrasonography, all of which are able to increase vessel perfusion and permeability^{247–249}; alternatively, pharmacological strategies, including agents targeting VEGFR signalling and those that generate nitric oxide, can normalize the tumour vasculature and thus promote the tumour-specific accumulation of nanomedicines^{203,250–252}. Furthermore, the addition of active targeting ligands, including antibodies, peptides and aptamers, provides a complementary strategy designed to increase the accumulation and retention of nanoparticles in tumours. However, despite a phase II clinical trial of BIND-014, a docetaxel-containing nanoparticle

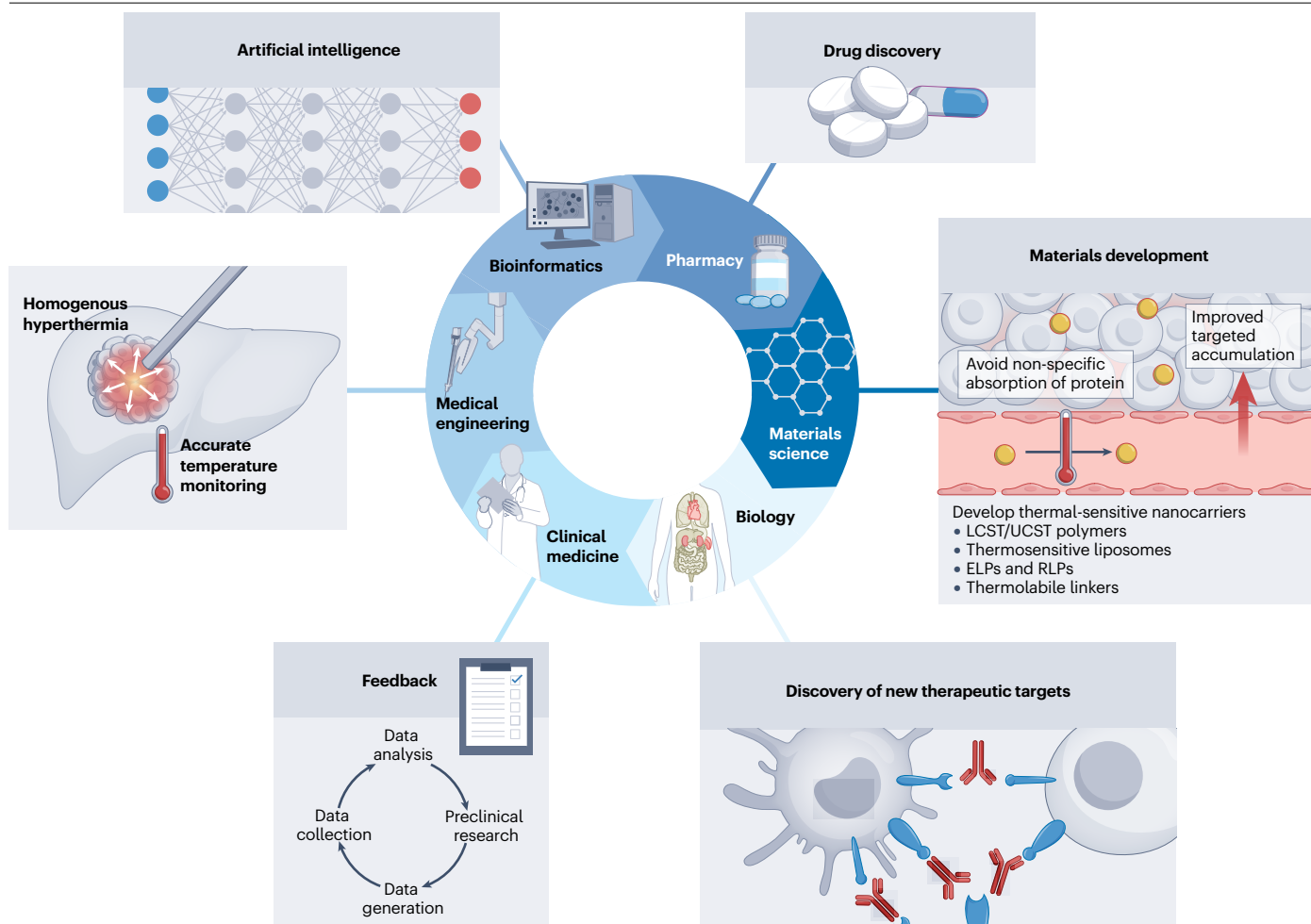


Fig. 4 | Future development of thermal immuno-nanomedicines driven by a multidisciplinary team. The clinical translation of thermal immuno-nanomedicines from promising preclinical candidates to approved therapies will require groups of experts, including materials scientists, biologists, oncologists, medical device engineers and even bioinformatics specialists to collaborate as

part of a multidisciplinary team to overcome the various challenges associated with the development of nanomedicines that combine the antitumour efficacy of hyperthermia and immunotherapy while minimizing the risks of adverse events. ELPs, elastin-like polypeptides; LCST, lower critical solution temperature; RLPs, resilin-like polypeptides; UCST, upper critical solution temperature.

targeted to prostate-specific membrane antigen (PSMA) in men with prostate cancer, no targeted nanomedicines have been approved for clinical use²⁵³. Several potential reasons exist for the largely disappointing results seen with nanomedicines in clinical trials; one of the major problems is the protein corona that forms around the surface of nanoparticles upon exposure to biological fluids^{254,255}. Nanoparticle surfaces are usually modified with hydrophilic polymers (such as PEG or zwitterionic ligands) to avoid interactions with the biological environment. Nonetheless, the non-specific absorption of proteins to form the protein corona has proven difficult to avoid, and this effect might mask surface-targeting ligands and/or trigger immunological recognition, leading to the rapid clearance of nanomedicines and/or adverse immune reactions.

Ultimately, outside of the current challenges associated with the use of nanomedicines, thermal therapies and/or immunotherapies, combining these modalities together in the same platform creates several new challenges. Firstly, although various TINs have been

prepared in labs, the scale-up of TIN production remains challenging. TIN preparation processes, including materials synthesis, encapsulation of therapeutic agents and product purification, typically lead to difficulties in reproducibility and quality control. Furthermore, combining different modalities not only potentially improves the capacity to kill cancer cells but might also increase the risk of adverse events. Hence, improving efficacy without a simultaneous increase in the incidence of adverse events is an important challenge. To achieve this goal, the dose ratio of different therapeutic agents, the power of thermal therapy devices, the choice of more biocompatible materials and the design of TINs must all be optimized to avoid overlapping toxicities and to maximize any synergistic therapeutic effects. Currently, most data on the performance of TINs are from preclinical studies involving mouse models. Therefore, these TINs should be tested in more complex animal models that better reflect the human immune system, which would enable the formulation of more effective TINs and the development of more precise treatment plans.

Conclusions

Overall, we conclude that developing more effective TINs requires collaboration among multidisciplinary teams of experts, including materials scientists, biologists, oncologists, medical device engineers and even bioinformatics specialists, to fully understand the mechanisms of action of hyperthermia-synergized immunotherapies, develop more effective combination platforms, and manufacture more effective heating and temperature monitoring devices. Addressing these points could further improve the performance of nanomedicines (Fig. 4). Various challenges currently impair both the development of more effective TINs and their clinical translation. Nonetheless, several reasons exist to anticipate that nanomedicines combining hyperthermia and immunotherapy will ultimately provide safe and effective cancer therapies.

Published online: 5 January 2023

References

- Nam, J. et al. Cancer nanomedicine for combination cancer immunotherapy. *Nat. Rev. Mater.* **4**, 398–414 (2019).
- Pearlman, A. H. et al. Targeting public neoantigens for cancer immunotherapy. *Nat. Cancer* **2**, 487–497 (2021).
- Li, Y. et al. Targeting IL-21 to tumor-reactive T cells enhances memory T cell responses and anti-PD-1 antibody therapy. *Nat. Commun.* **12**, 951 (2021).
- Sahin, U. et al. An RNA vaccine drives immunity in checkpoint-inhibitor-treated melanoma. *Nature* **585**, 107–112 (2020).
- Waldman, A. D., Fritz, J. M. & Lenardo, M. J. A guide to cancer immunotherapy: from T cell basic science to clinical practice. *Nat. Rev. Immunol.* **20**, 651–668 (2020).
- Singh, A. K. & McGuirk, J. P. CAR T cells: continuation in a revolution of immunotherapy. *Lancet Oncol.* **21**, E168–E178 (2020).
- Vitale, I., Shema, E., Loi, S. & Galluzzi, L. Intratumoral heterogeneity in cancer progression and response to immunotherapy. *Nat. Med.* **27**, 212–224 (2021).
- Newman, J. H. et al. Intratumoral injection of the seasonal flu shot converts immunologically cold tumors to hot and serves as an immunotherapy for cancer. *Proc. Natl Acad. Sci. USA* **117**, 1119–1128 (2020).
- Jackson, C. M., Choi, J. & Lim, M. Mechanisms of immunotherapy resistance: lessons from glioblastoma. *Nat. Immunol.* **20**, 1100–1109 (2019).
- Bonaventura, P. et al. Cold tumors: a therapeutic challenge for immunotherapy. *Front. Immunol.* **10**, 168 (2019).
- Certo, M., Tsai, C. H., Pucino, V., Ho, P. C. & Mauro, C. Lactate modulation of immune responses in inflammatory versus tumour microenvironments. *Nat. Rev. Immunol.* **21**, 151–161 (2021).
- Chen, Y., McAndrews, K. M. & Kalluri, R. Clinical and therapeutic relevance of cancer-associated fibroblasts. *Nat. Rev. Clin. Oncol.* **18**, 792–804 (2021).
- Rodriguez Garcia, A. et al. CAR-T cell-mediated depletion of immunosuppressive tumor-associated macrophages promotes endogenous antitumor immunity and augments adoptive immunotherapy. *Nat. Commun.* **12**, 877 (2021).
- Schmid, P. et al. Atezolizumab plus nab-paclitaxel as first-line treatment for unresectable, locally advanced or metastatic triple-negative breast cancer (IMpassion130): updated efficacy results from a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol.* **21**, 44–59 (2020).
- Adams, S. et al. Atezolizumab plus nab-paclitaxel in the treatment of metastatic triple-negative breast cancer with 2-year survival follow-up a phase 1b clinical trial. *JAMA Oncol.* **5**, 334–342 (2019).
- Esperanza Rodriguez Ruiz, M., Vitale, I., Harrington, K. J., Melero, I. & Galluzzi, L. Immunological impact of cell death signaling driven by radiation on the tumor microenvironment. *Nat. Immunol.* **21**, 120–134 (2020).
- Patel, R. B. et al. Low-dose targeted radionuclide therapy renders immunologically cold tumors responsive to immune checkpoint blockade. *Sci. Transl. Med.* **13**, eabb3631 (2021).
- Chang, M., Hou, Z., Wang, M., Li, C. & Lin, J. Recent advances in hyperthermia therapy-based synergistic immunotherapy. *Adv. Mater.* **33**, 2004788 (2021).
- Cortes, J. et al. Pembrolizumab plus chemotherapy versus placebo plus chemotherapy for previously untreated locally recurrent inoperable or metastatic triple-negative breast cancer (KEYNOTE-355): a randomised, placebo-controlled, double-blind, phase 3 clinical trial. *Lancet* **396**, 1817–1828 (2020).
- Schmid, P. et al. Pembrolizumab for early triple-negative breast cancer. *N. Engl. J. Med.* **382**, 810–821 (2020).
- Liu, W. et al. Combination of immune checkpoint inhibitors with chemotherapy in lung cancer. *OncoTargets Ther.* **13**, 7229–7241 (2020).
- Larroquette, M. et al. Combining immune checkpoint inhibitors with chemotherapy in advanced solid tumours: a review. *Eur. J. Cancer* **158**, 47–62 (2021).
- Zhou, J. L., Huang, Q., Huang, Z. J. & Li, J. Q. Combining immunotherapy and radiotherapy in lung cancer: a promising future? *J. Thorac. Dis.* **12**, 4498–4503 (2020).
- Shirvalilou, S. et al. Magnetic hyperthermia as an adjuvant cancer therapy in combination with radiotherapy versus radiotherapy alone for recurrent/progressive glioblastoma: a systematic review. *J. Neurooncol.* **152**, 419–428 (2021).
- Li, Q. et al. Pre- and post-irradiation mild hyperthermia enabled by NIR-II for sensitizing radiotherapy. *Biomaterials* **257**, 120235 (2020).
- Sen, A. et al. Mild elevation of body temperature reduces tumor interstitial fluid pressure and hypoxia and enhances efficacy of radiotherapy in murine tumor models. *Cancer Res.* **71**, 3872–3880 (2011).
- Mu, C. et al. Chemotherapy sensitizes therapy-resistant cells to mild hyperthermia by suppressing heat shock protein 27 expression in triple-negative breast cancer. *Clin. Cancer Res.* **24**, 4900–4912 (2018).
- Klaver, C. E. L. et al. Adjuvant hyperthermic intraperitoneal chemotherapy in patients with locally advanced colon cancer (COLOPEC): a multicentre, open-label, randomised trial. *Lancet Gastroenterol.* **4**, 761–770 (2019).
- Shin, D. H., Melnick, K. F., Tran, D. D. & Ghiaieddin, A. P. In situ vaccination with laser interstitial thermal therapy augments immunotherapy in malignant gliomas. *J. Neurooncol.* **151**, 85–92 (2021).
- Napoli, A. et al. Noninvasive therapy for osteoid osteoma: a prospective developmental study with MR imaging-guided high-intensity focused ultrasound. *Radiology* **285**, 186–196 (2017).
- Friedman, M. et al. Radiofrequency ablation of cancer. *Cardiovasc. Interv. Radiol.* **27**, 427–434 (2004).
- Jiang, Y., Huang, J., Xu, C. & Pu, K. Activatable polymer nanoaggonist for second near-infrared photothermal immunotherapy of cancer. *Nat. Commun.* **12**, 742 (2021).
- Fite, B. Z. et al. Immune modulation resulting from MR-guided high intensity focused ultrasound in a model of murine breast cancer. *Sci. Rep.* **11**, 927 (2021).
- Pan, J. et al. Combined magnetic hyperthermia and Immune therapy for primary and metastatic tumor treatments. *ACS Nano* **14**, 1033–1044 (2020).
- Lerner, E. C., Edwards, R. M., Wilkinson, D. S. & Fecci, P. E. Laser ablation: heating up the anti-tumor response in the intracranial compartment. *Adv. Drug Deliv. Rev.* **185**, 114311 (2022).
- Pan, Y., Liu, L., Rao, L. & Chen, X. Nanomaterial-mediated ablation therapy for cancer stem cells. *Matter* **5**, 1367–1390 (2022).
- Pan, J., Xu, Y., Wu, Q., Hu, P. & Shi, J. Mild magnetic hyperthermia-activated innate immunity for liver cancer therapy. *J. Am. Chem. Soc.* **143**, 8116–8128 (2021).
- Chen, Q. et al. Fever-range thermal stress promotes lymphocyte trafficking across high endothelial venules via an interleukin 6 trans-signaling mechanism. *Nat. Immunol.* **7**, 1299–1308 (2006).
- Shi, L. et al. Thermal ablation plus toripalimab in patients with advanced hepatocellular carcinoma: phase I results from a multicenter, open-label, controlled phase I/II trial (IRI1330). *Ann. Oncol.* **32**, S826–S826 (2021).
- Lyu, N. et al. Ablation reboots the response in advanced hepatocellular carcinoma with stable or atypical response during PD-1 therapy: a proof-of-concept study. *Front. Oncol.* **10**, 580241 (2020).
- Duffy, A. G. et al. Tremelimumab in combination with ablation in patients with advanced hepatocellular carcinoma. *J. Hepatol.* **66**, 545–551 (2017).
- Xie, C. et al. Tremelimumab in combination with microwave ablation in patients with refractory biliary tract cancer. *Hepatology* **69**, 2048–2060 (2019).
- Zuo, S. et al. Nano-immunotherapy for each stage of cancer cellular immunity: which, why, and what? *Theranostics* **11**, 7471–7487 (2021).
- Bailey, S. R. & Maus, M. V. Gene editing for immune cell therapies. *Nat. Biotechnol.* **37**, 1425–1434 (2019).
- Fierro, J. et al. Dual-sgRNA CRISPR/Cas9 knockout of PD-L1 in human U87 glioblastoma tumor cells inhibits proliferation, invasion, and tumor-associated macrophage polarization. *Sci. Rep.* **12**, 2417 (2022).
- Shi, L. et al. CRISPR knock out CTLA-4 enhances the anti-tumor activity of cytotoxic T lymphocytes. *Gene* **636**, 36–41 (2017).
- Chen, Q. et al. Photothermal therapy with immune-adjuvant nanoparticles together with checkpoint blockade for effective cancer immunotherapy. *Nat. Commun.* **7**, 13193 (2016).
- Huang, L. et al. Mild photothermal therapy potentiates anti-PD-L1 treatment for immunologically cold tumors via an all-in-one and all-in-control strategy. *Nat. Commun.* **10**, 4871 (2019).
- Saccomandi, P., Lapergola, A., Longo, F., Schena, E. & Quero, G. Thermal ablation of pancreatic cancer: a systematic literature review of clinical practice and pre-clinical studies. *Int. J. Hyperth.* **35**, 398–418 (2019).
- Chu, K. F. & Dupuy, D. E. Thermal ablation of tumours: biological mechanisms and advances in therapy. *Nat. Rev. Cancer* **14**, 199–208 (2014).
- van Solinge, T. S., Nieland, L., Chiocia, E. A. & Broekman, M. L. D. Advances in local therapy for glioblastoma — taking the fight to the tumour. *Nat. Rev. Neurol.* **18**, 221–236 (2022).
- Elming, P. B. et al. Hyperthermia: the optimal treatment to overcome radiation resistant hypoxia. *Cancers* **11**, 60 (2019).
- Issels, R. D. et al. Effect of neoadjuvant chemotherapy plus regional hyperthermia on long-term outcomes among patients with localized high-risk soft tissue sarcoma. *JAMA Oncol.* **4**, 483–492 (2018).
- Krawczyk, P. M. et al. Mild hyperthermia inhibits homologous recombination, induces BRCA2 degradation, and sensitizes cancer cells to poly (ADP-ribose) polymerase-1 inhibition. *Proc. Natl Acad. Sci. USA* **108**, 9851–9856 (2011).

55. Bailey, C. W. & Sydnor, M. K. Current state of tumor ablation therapies. *Dig. Dis. Sci.* **64**, 951–958 (2019).
56. Belyanchikov, M. A. et al. Dielectric ordering of water molecules arranged in a dipolar lattice. *Nat. Commun.* **11**, 3927 (2020).
57. Kok, H. P. et al. Heating technology for malignant tumors: a review. *Int. J. Hyperth.* **37**, 711–741 (2020).
58. Gao, D. et al. NIR/MRI-guided oxygen-independent carrier-free anti-tumor nano-theranostics. *Small* **18**, 2106000 (2021).
59. Gao, D. et al. Multifunctional phototheranostic nanomedicine for cancer imaging and treatment. *Mater. Today Bio* **5**, 100035 (2020).
60. Gao, D. et al. Targeting hypoxic tumors with hybrid nanobullets for oxygen-independent synergistic photothermal and thermodynamic therapy. *Nanomicro. Lett.* **13**, 99 (2021).
61. Hammill, C. W. et al. Evaluation of a minimally invasive image-guided surgery system for hepatic ablation procedures. *Surg. Innov.* **21**, 419–426 (2014).
62. Patel, P., Patel, N. V. & Danish, S. F. Intracranial MR-guided laser-induced thermal therapy: single-center experience with the Visualase thermal therapy system. *J. Neurosurg.* **125**, 853–860 (2016).
63. Mohammadi, A. M. & Schroeder, J. L. Laser interstitial thermal therapy in treatment of brain tumors — the NeuroBlate system. *Expert Rev. Med. Devices* **11**, 109–119 (2014).
64. Sloan, A. E. et al. Results of the neuroblate system first-in-humans phase I clinical trial for recurrent glioblastoma. *J. Neurosurg.* **118**, 1202–1219 (2013).
65. Li, X. S., Lovell, J. F., Yoon, J. & Chen, X. Y. Clinical development and potential of photothermal and photodynamic therapies for cancer. *Nat. Rev. Clin. Oncol.* **17**, 657–674 (2020).
66. Xia, Y. et al. Long-term effects of repeat hepatectomy vs percutaneous radiofrequency ablation among patients with recurrent hepatocellular carcinoma. *JAMA Oncol.* **6**, 255–263 (2020).
67. Kinoshita, T. RFA experiences, indications and clinical outcomes. *Int. J. Clin. Oncol.* **24**, 603–607 (2019).
68. Al Zubaidi, M., Lotter, K., Marshall, M. & Lozinskiy, M. Radiofrequency ablation for renal tumours: a retrospective study from a tertiary centre. *Asian J. Urol.* **9**, 2214–2219 (2021).
69. Wan, J. & Wang, J. Current progression of radiofrequency ablation (RFA) in clinical application of lung cancer therapy. *J. Biomater. Tissue Eng.* **9**, 417–426 (2019).
70. Vietti Violi, N. et al. Efficacy of microwave ablation versus radiofrequency ablation for the treatment of hepatocellular carcinoma in patients with chronic liver disease: a randomised controlled phase 2 trial. *Lancet Gastroenterol. Hepatol.* **3**, 317–325 (2018).
71. Schirmang, T. C. & Dupuy, D. E. Image-guided thermal ablation of nonresectable hepatic tumors using the Cool-Tip radiofrequency ablation system. *Expert Rev. Med. Devices* **4**, 803–814 (2007).
72. Gaffke, G. et al. Use of semiflexible applicators for radiofrequency ablation of liver tumors. *Cardiovasc. Interv. Radiol.* **29**, 270–275 (2006).
73. Solmi, L., Nigro, G. & Roda, E. Therapeutic effectiveness of echo-guided percutaneous radiofrequency ablation therapy with a LeVeen needle electrode in hepatocellular carcinoma. *World J. Gastroenterol.* **12**, 1098–1104 (2006).
74. Mohkam, K. et al. No-touch multipolar radiofrequency ablation vs. surgical resection for solitary hepatocellular carcinoma ranging from 2 to 5 cm. *J. Hepatol.* **68**, 1172–1180 (2018).
75. King, J. et al. Percutaneous radiofrequency ablation of pulmonary metastases in patients with colorectal cancer. *Br. J. Surg.* **91**, 217–223 (2004).
76. Sommer, C. M. et al. CT-guided bipolar and multipolar radiofrequency ablation (RF ablation) of renal cell carcinoma: Specific technical aspects and clinical results. *Cardiovasc. Interv. Radiol.* **36**, 731–737 (2013).
77. Wells, C. D., Kim, H. J., Moirano, M. M., Fleischer, D. E. & Sharma, V. K. Successful ablation of Barrett esophagus (BE) with dysplasia using the Halo360 ablation system: a single-center experience. *Am. J. Gastroenterol.* **101**, S535–S535 (2006).
78. Rothstein, R. I., Chang, K. J., Overholt, B. F., Bergman, J. J. & Shaheen, N. J. Focal ablation for treatment of dysplastic and non-dysplastic Barrett esophagus: safety profile and initial experience with the Halo90 device in 508 cases. *Gastrointest. Endosc.* **65**, AB147–AB147 (2007).
79. Shen, R. et al. Ultrasonography-guided radiofrequency ablation combined with lauracemacrogol sclerotherapy for mixed thyroid nodules. *Am. J. Transl. Res.* **13**, 5035–5042 (2021).
80. Mayer, T. et al. Spinal metastases treated with bipolar radiofrequency ablation with increased (>70°C) target temperature: pain management and local tumor control. *Diagn. Interv. Imaging* **102**, 27–34 (2021).
81. Lorenc, T., Kocon, H. & Golebiowski, M. Computed tomography-guided percutaneous radiofrequency and laser ablation for the treatment of osteoid osteoma — long-term follow-up from 5 to 10 years. *Pol. J. Radiol.* **86**, E19–E30 (2021).
82. Osaki, Y. et al. Clinical effectiveness of bipolar radiofrequency ablation for small liver cancers. *J. Gastroenterol.* **48**, 874–883 (2013).
83. Llovet, J. M. et al. Locoregional therapies in the era of molecular and immune treatments for hepatocellular carcinoma. *Nat. Rev. Gastroenterol. Hepatol.* **18**, 293–313 (2021).
84. Yu, M. et al. Microwave ablation of primary breast cancer inhibits metastatic progression in model mice via activation of natural killer cells. *Cell. Mol. Immunol.* **18**, 2153–2164 (2021).
85. Vogl, T. J. et al. A comparison between 915 MHz and 2450 MHz microwave ablation systems for the treatment of small diameter lung metastases. *Diagn. Interv. Radiol.* **24**, 31–37 (2018).
86. Livraghi, T., Meloni, F., Solbiati, L. & Zanusi, G. Complications of microwave ablation for liver tumors: results of a multicenter study. *Cardiovasc. Interv. Radiol.* **35**, 868–874 (2012).
87. Imajo, K. et al. New microwave ablation system for unresectable liver tumors that forms large, spherical ablation zones. *J. Gastroenterol. Hepatol.* **33**, 2007–2014 (2018).
88. Alexander, E. S. et al. Microwave ablation of focal hepatic malignancies regardless of size: a 9-year retrospective study of 64 patients. *Eur. J. Radiol.* **84**, 1083–1090 (2015).
89. Habert, P. et al. Percutaneous lung and liver CT-guided ablation on swine model using microwave ablation to determine ablation size for clinical practice. *Int. J. Hyperth.* **38**, 1140–1148 (2021).
90. Medhat, E. et al. Value of microwave ablation in treatment of large lesions of hepatocellular carcinoma. *J. Dig. Dis.* **16**, 456–463 (2015).
91. Filippiadis, D. K. et al. Computed tomography-guided percutaneous microwave ablation of hepatocellular carcinoma in challenging locations: safety and efficacy of high-power microwave platforms. *Int. J. Hyperth.* **34**, 863–869 (2018).
92. Zhao, H. & Steinke, K. Long-term outcome following microwave ablation of early-stage non-small cell lung cancer. *J. Med. Imaging Radiat. Oncol.* **64**, 787–793 (2020).
93. Pusceddu, C., Sotgia, B., Fele, R. M. & Melis, L. Treatment of bone metastases with microwave thermal ablation. *J. Vasc. Interv. Radiol.* **24**, 229–233 (2013).
94. Rostas, J. W., Hong, Y. K., Schulz, B., Sanders, M. E. & Martin, R. C. G. A multi-visceral study comparing three proprietary microwave ablation devices. *HPB* **19**, S158–S159 (2017).
95. Peek, M. C. L. et al. Minimally invasive ablative techniques in the treatment of breast cancer: a systematic review and meta-analysis. *Int. J. Hyperth.* **33**, 191–202 (2017).
96. Lindner, U. et al. Focal magnetic resonance guided focused ultrasound for prostate cancer: initial North American experience. *Can. Urol. Assoc. J.* **6**, E283–E286 (2012).
97. Francica, G. Needle track seeding after radiofrequency ablation for hepatocellular carcinoma: prevalence, impact, and management challenge. *J. Hepatocell. Carcinoma* **4**, 23–27 (2017).
98. Kennedy, J. E. High-intensity focused ultrasound in the treatment of solid tumours. *Nat. Rev. Cancer* **5**, 321–327 (2005).
99. Rebillard, X. et al. Transrectal high-intensity focused ultrasound in the treatment of localized prostate cancer. *J. Endourol.* **19**, 693–701 (2005).
100. Aliaev, I. et al. Treatment of prostatic cancer with high intensity focused ultrasound (HIFU) using Ablatherm device. *Urologia* **6**, 39–44 (2007).
101. Burtnyk, M., Hill, T., Cadieux Pitre, H. & Welch, I. Magnetic resonance image guided transurethral ultrasound prostate ablation: a preclinical safety and feasibility study with 28-day followup. *J. Urol.* **193**, 1669–1675 (2015).
102. Meng, Y., Hynynen, K. & Lipsman, N. Applications of focused ultrasound in the brain: from thermoablation to drug delivery. *Nat. Rev. Neurol.* **17**, 7–22 (2021).
103. Temple, M. J. et al. Establishing a clinical service for the treatment of osteoid osteoma using magnetic resonance-guided focused ultrasound: overview and guidelines. *J. Ther. Ultrasound* **4**, 16–16 (2016).
104. Dobrotwir, A. & Pun, E. Clinical 24 month experience of the first MRgFUS Unit for treatment of uterine fibroids in Australia. *J. Med. Imaging Radiat. Oncol.* **56**, 409–416 (2012).
105. Knüttel, F. M. et al. Early health technology assessment of magnetic resonance-guided high intensity focused ultrasound ablation for the treatment of early-stage breast cancer. *J. Ther. Ultrasound* **5**, 1–10 (2017).
106. Najafi, A., Fuchs, B. & Binkert, C. A. Mid-term results of MR-guided high-intensity focused ultrasound treatment for relapsing superficial desmoids. *Int. J. Hyperth.* **36**, 537–541 (2019).
107. Jung, S. E., Cho, S. H., Jang, J. H. & Han, J. Y. High-intensity focused ultrasound ablation in hepatic and pancreatic cancer: complications. *Abdom. Imaging* **36**, 185–195 (2011).
108. Zhao, J. et al. The efficacy of a new high intensity focused ultrasound therapy for locally advanced pancreatic cancer. *J. Cancer Res. Clin. Oncol.* **143**, 2105–2111 (2017).
109. Leslie, T. A. et al. High intensity focused ultrasound in the treatment of small kidney tumours — the Oxford experience. *J. Clin. Oncol.* **24**, 14645–14645 (2006).
110. Arcot, R. & Polascik, T. J. Does MRI-guided TULSA provide a targeted approach to ablation? *Nat. Rev. Urol.* **18**, 5–6 (2021).
111. Nair, S. M. et al. Magnetic resonance imaging-guided transurethral ultrasound ablation in patients with localised prostate cancer: 3-year outcomes of a prospective phase I study. *BJU Int.* **127**, 544–552 (2021).
112. Muto, S. et al. Focal therapy with high-intensity-focused ultrasound in the treatment of localized prostate cancer. *Jpn. J. Clin. Oncol.* **38**, 192–199 (2008).
113. Gilchrist, R. K. et al. Selective inductive heating of lymph nodes. *Ann. Surg.* **146**, 596–606 (1957).
114. Gavilan, H. et al. Magnetic nanoparticles and clusters for magnetic hyperthermia: optimizing their heat performance and developing combinatorial therapies to tackle cancer. *Chem. Soc. Rev.* **50**, 11614–11667 (2021).
115. Song, G. et al. Carbon-coated FeCo nanoparticles as sensitive magnetic-particle-imaging tracers with photothermal and magnetothermal properties. *Nat. Biomed. Eng.* **4**, 325–334 (2020).
116. Liu, X. et al. Comprehensive understanding of magnetic hyperthermia for improving antitumor therapeutic efficacy. *Theranostics* **10**, 3793–3815 (2020).
117. Lu, C. et al. Engineering of magnetic nanoparticles as magnetic particle imaging tracers. *Chem. Soc. Rev.* **50**, 8102–8146 (2021).
118. Ho, D., Sun, X. & Sun, S. Monodisperse magnetic nanoparticles for theranostic applications. *Acc. Chem. Res.* **44**, 875–882 (2011).

119. Maier-Hauff, K. et al. Efficacy and safety of intratumoral thermotherapy using magnetic iron-oxide nanoparticles combined with external beam radiotherapy on patients with recurrent glioblastoma multiforme. *J. Neurooncol.* **103**, 317–324 (2011).
120. Galluzzi, L., Buque, A., Kepp, O., Zitvogel, L. & Kroemer, G. Immunogenic cell death in cancer and infectious disease. *Nat. Rev. Immunol.* **17**, 97–111 (2017).
121. Garg, A. D. & Agostinis, P. Cell death and immunity in cancer: from danger signals to mimicry of pathogen defense responses. *Immunol. Rev.* **280**, 126–148 (2017).
122. Li, Z., Deng, J., Sun, J. & Ma, Y. Hyperthermia targeting the tumor microenvironment facilitates immune checkpoint inhibitors. *Front. Immunol.* **11**, 595207 (2020).
123. Krysko, D. V. et al. Immunogenic cell death and DAMPs in cancer therapy. *Nat. Rev. Cancer* **12**, 860–875 (2012).
124. Munoz, N. M. et al. Immune modulation by molecularly targeted photothermal ablation in a mouse model of advanced hepatocellular carcinoma and cirrhosis. *Sci. Rep.* **12**, 14449 (2022).
125. Guo, D. F. et al. Exosomes from heat-stressed tumour cells inhibit tumour growth by converting regulatory T cells to Th17 cells via IL-6. *Immunology* **154**, 132–143 (2018).
126. Mace, T. A., Zhong, L. W., Kokolus, K. M. & Repasky, E. A. Effector CD8⁺ T cell IFN- γ production and cytotoxicity are enhanced by mild hyperthermia. *Int. J. Hyperther.* **28**, 9–18 (2012).
127. Chen, T., Guo, J., Han, C., Yang, M. & Cao, X. Heat shock protein 70, released from heat-stressed tumor cells, initiates antitumor immunity by inducing tumor cell chemokine production and activating dendritic cells via TLR4 pathway. *J. Immunol.* **182**, 1449–1459 (2009).
128. Zhao, W. et al. Hyperthermia differentially regulates TLR4 and TLR2-mediated innate immune response. *Immunol. Lett.* **108**, 137–142 (2007).
129. Gallucci, S., Lolkema, M. & Matzinger, P. Natural adjuvants: endogenous activators of dendritic cells. *Nat. Med.* **5**, 1249–1255 (1999).
130. Ostberg, J. R., Dayanc, B. E., Yuan, M., Oflazoglu, E. & Repasky, E. A. Enhancement of natural killer (NK) cell cytotoxicity by fever-range thermal stress is dependent on NKG2D function and is associated with plasma membrane NKG2D clustering and increased expression of MICA on target cells. *J. Leukoc. Biol.* **82**, 1322–1331 (2007).
131. Bae, J.-H. et al. Quercetin enhances susceptibility to NK cell-mediated lysis of tumor cells through induction of NKG2D ligands and suppression of HSP70. *J. Immunother.* **33**, 391–401 (2010).
132. He, K., Liu, P. & Xu, L. X. The cryo-thermal therapy eradicated melanoma in mice by eliciting CD4⁺ T-cell-mediated antitumor memory immune response. *Cell Death Dis.* **8**, e2703 (2017).
133. Liu, Y. et al. Plasmonic gold nanostar-mediated photothermal immunotherapy for brain tumor ablation and immunologic memory. *Immunother* **11**, 1293–1302 (2019).
134. Hurwitz, M. D. Hyperthermia and immunotherapy: clinical opportunities. *Int. J. Hyperther.* **36**, 4–9 (2019).
135. Rittmeyer, A. et al. Atezolizumab versus docetaxel in patients with previously treated non-small-cell lung cancer (OAK): a phase 3, open-label, multicentre randomised controlled trial. *Lancet* **389**, 255–265 (2017).
136. Wang, S. H. et al. Nanoparticle-based medicines in clinical cancer therapy. *Nano Today* **45**, 101512 (2022).
137. Cortes, J. et al. Pembrolizumab plus chemotherapy in advanced triple-negative breast cancer. *N. Engl. J. Med.* **387**, 217–226 (2022).
138. Paz Ares, L. et al. Pembrolizumab plus chemotherapy for squamous non-small-cell lung cancer. *N. Engl. J. Med.* **379**, 2040–2051 (2018).
139. Yang, Z. et al. Fighting immune cold and reprogramming immunosuppressive tumor microenvironment with red blood cell membrane-camouflaged nanobullets. *ACS Nano* **14**, 17442–17457 (2020).
140. Chaudhary, N., Weissman, D. & Whitehead, K. A. mRNA vaccines for infectious diseases: principles, delivery and clinical translation. *Nat. Rev. Drug Discov.* **20**, 817–838 (2021).
141. Saxena, M., van der Burg, S. H., Melief, C. J. M. & Bhardwaj, N. Therapeutic cancer vaccines. *Nat. Rev. Cancer* **21**, 360–378 (2021).
142. Teijaro, J. R. & Farber, D. L. COVID-19 vaccines: modes of immune activation and future challenges. *Nat. Rev. Immunol.* **21**, 195–197 (2021).
143. Li, A. W. et al. A facile approach to enhance antigen response for personalized cancer vaccination. *Nat. Mater.* **17**, 528–534 (2018).
144. Kuai, R., Ochyl, L. J., Bahjat, K. S., Schwendeman, A. & Moon, J. J. Designer vaccine nanodiscs for personalized cancer immunotherapy. *Nat. Mater.* **16**, 489–496 (2017).
145. Aikins, M. E., Xu, C. & Moon, J. J. Engineered nanoparticles for cancer vaccination and immunotherapy. *Acc. Chem. Res.* **53**, 2094–2105 (2020).
146. Chen, P. M. et al. Modulation of tumor microenvironment using a TLR-7/8 agonist-loaded nanoparticle system that exerts low-temperature hyperthermia and immunotherapy for in situ cancer vaccination. *Biomaterials* **230**, 119629 (2020).
147. Liu, L. et al. Cell membrane coating integrity affects the internalization mechanism of biomimetic nanoparticles. *Nat. Commun.* **12**, 5726 (2021).
148. Zhou, J. R., Kroll, A. V., Holay, M., Fang, R. H. & Zhang, L. F. Biomimetic nanotechnology toward personalized vaccines. *Adv. Mater.* **32**, 1901255 (2020).
149. Liu, W. L. et al. Cytomembrane nanovaccines show therapeutic effects by mimicking tumor cells and antigen presenting cells. *Nat. Commun.* **10**, 3199 (2019).
150. Chen, Q. et al. A hybrid eukaryotic-prokaryotic nanoplatfrom with photothermal modality for enhanced antitumor vaccination. *Adv. Mater.* **32**, 1908185 (2020).
151. Wang, T. et al. A cancer vaccine-mediated postoperative immunotherapy for recurrent and metastatic tumors. *Nat. Commun.* **9**, 1532 (2018).
152. Sharma, P. & Allison, J. P. The future of immune checkpoint therapy. *Science* **348**, 56–61 (2015).
153. Kubli, S. P., Berger, T., Araujo, D. V., Siu, L. L. & Mak, T. W. Beyond immune checkpoint blockade: emerging immunological strategies. *Nat. Rev. Drug Discov.* **20**, 899–919 (2021).
154. Mellman, I., Coukos, G. & Dranoff, G. Cancer immunotherapy comes of age. *Nature* **480**, 480–489 (2011).
155. Lin, D. Y. W. et al. The PD-1/PD-L1 complex resembles the antigen-binding Fv domains of antibodies and T cell receptors. *Proc. Natl Acad. Sci. USA* **105**, 3011–3016 (2008).
156. Leach, D. R., Krummel, M. F. & Allison, J. P. Enhancement of antitumor immunity by CTLA-4 blockade. *Science* **271**, 1734–1736 (1996).
157. Pallotta, M. T. et al. Indoleamine 2,3-dioxygenase is a signaling protein in long-term tolerance by dendritic cells. *Nat. Immunol.* **12**, 870–878 (2011).
158. Dong, C. et al. ICOS co-stimulatory receptor is essential for T-cell activation and function. *Nature* **409**, 97–101 (2001).
159. Marangoni, F. et al. Expansion of tumor-associated Treg cells upon disruption of a CTLA-4-dependent feedback loop. *Cell* **184**, 3998–4015 (2021).
160. Zaidi, N. & Jaffee, E. M. Immunotherapy transforms cancer treatment. *J. Clin. Invest.* **129**, 46–47 (2019).
161. Wang, Z. et al. Janus nanobullets combine photodynamic therapy and magnetic hyperthermia to potentiate synergetic anti-metastatic immunotherapy. *Adv. Sci.* **6**, 1901690 (2019).
162. Liu, J. et al. Tumor hypoxia-activated combinatorial nanomedicine triggers systemic antitumor immunity to effectively eradicate advanced breast cancer. *Biomaterials* **273**, 120847 (2021).
163. Wang, C. et al. Immunological responses triggered by photothermal therapy with carbon nanotubes in combination with anti-CTLA-4 therapy to inhibit cancer metastasis. *Adv. Mater.* **26**, 8154–8162 (2014).
164. Chao, Y. et al. Iron nanoparticles for low-power local magnetic hyperthermia in combination with immune checkpoint blockade for systemic antitumor therapy. *Nano Lett.* **19**, 4287–4296 (2019).
165. Patsoukis, N., Wang, Q., Strauss, L. & Boussiotis Vassiliki, A. Revisiting the PD-1 pathway. *Sci. Adv.* **6**, eabd2712 (2020).
166. Chen, L. & Han, X. Anti-PD-1/PD-L1 therapy of human cancer: past, present, and future. *J. Clin. Invest.* **125**, 3384–3391 (2015).
167. Li, H., van der Merwe, P. A. & Sivakumar, S. Biomarkers of response to PD-1 pathway blockade. *Br. J. Cancer* **126**, 1663–1675 (2022).
168. Liu, X. et al. Ferrimagnetic vortex nanoring-mediated mild magnetic hyperthermia imparts potent immunological effect for treating cancer metastasis. *ACS Nano* **13**, 8811–8825 (2019).
169. Zhang, Y. et al. Native mitochondria-targeting polymeric nanoparticles for mild photothermal therapy rationally potentiated with immune checkpoints blockade to inhibit tumor recurrence and metastasis. *Chem. Eng. J.* **424**, 130171 (2021).
170. Duan, X., Chan, C. & Lin, W. Nanoparticle-mediated immunogenic cell death enables and potentiates cancer immunotherapy. *Angew. Chem. Int. Ed.* **58**, 670–680 (2019).
171. Martins, F. et al. Adverse effects of immune-checkpoint inhibitors: epidemiology, management and surveillance. *Nat. Rev. Clin. Oncol.* **16**, 563–580 (2019).
172. McManus, M. T. & Sharp, P. A. Gene silencing in mammals by small interfering RNAs. *Nat. Rev. Genet.* **3**, 737–747 (2002).
173. Saha, K. et al. The NIH somatic cell genome editing program. *Nature* **592**, 195–204 (2021).
174. Kinc, A. et al. The Onpatro story and the clinical translation of nanomedicines containing nucleic acid-based drugs. *Nat. Nanotechnol.* **14**, 1084–1087 (2019).
175. Saw, P. E. & Song, E. W. siRNA therapeutics: a clinical reality. *Sci. China Life Sci.* **63**, 485–500 (2020).
176. Liu, B. et al. Effects of gold nanoprisms-assisted human PD-L1 siRNA on both gene down-regulation and photothermal therapy on lung cancer. *Acta Biomater.* **99**, 307–319 (2019).
177. Sánchez Rivera, F. J. & Jacks, T. Applications of the CRISPR–Cas9 system in cancer biology. *Nat. Rev. Cancer* **15**, 387–393 (2015).
178. Grunwald, H. A., Weitzel, A. J. & Cooper, K. L. Applications of and considerations for using CRISPR–Cas9-mediated gene conversion systems in rodents. *Nat. Protoc.* **17**, 3–14 (2022).
179. Strzyp, P. CRISPR–Cas9 wins nobel. *Nat. Rev. Mol. Cell Biol.* **21**, 714–714 (2020).
180. Wang, D., Tai, P. W. L. & Gao, G. Adeno-associated virus vector as a platform for gene therapy delivery. *Nat. Rev. Drug Discov.* **18**, 358–378 (2019).
181. Bulcha, J. T., Wang, Y., Ma, H., Tai, P. W. L. & Gao, G. Viral vector platforms within the gene therapy landscape. *Signal Transduct. Target. Ther.* **6**, 53 (2021).
182. Ou, X., Ma, Q., Yin, W., Ma, X. & He, Z. CRISPR/Cas9 gene-editing in cancer immunotherapy: promoting the present revolution in cancer therapy and exploring more. *Front. Cell Dev. Biol.* **9**, 674467 (2021).
183. Gillmore, J. D. et al. CRISPR–Cas9 in vivo gene editing for transthyretin amyloidosis. *N. Engl. J. Med.* **385**, 493–502 (2021).
184. Tang, H. et al. Reprogramming the tumor microenvironment through second-near-infrared-window photothermal genome editing of PD-L1 mediated by supramolecular gold nanorods for enhanced cancer immunotherapy. *Adv. Mater.* **33**, 2006003 (2021).
185. Logtenberg, M. E. W., Scheeren, F. A. & Schumacher, T. N. The CD47–SIRP α immune checkpoint. *Immunity* **52**, 742–752 (2020).
186. Sikic, B. I. et al. First-in-human, first-in-class phase I trial of the anti-CD47 antibody Hu5F9-G4 in patients with advanced cancers. *J. Clin. Oncol.* **37**, 946–953 (2019).

187. Advani, R. et al. CD47 blockade by Hu5F9-G4 and Rituximab in non-hodgkin's lymphoma. *N. Engl. J. Med.* **379**, 1711–1721 (2018).
188. Liu, X. et al. CD47 blockade triggers T cell-mediated destruction of immunogenic tumors. *Nat. Med.* **21**, 1209–1215 (2015).
189. Zhou, F. et al. Tumor microenvironment-activatable prodrug vesicles for nanoenabled cancer chemoimmunotherapy combining immunogenic cell death induction and CD47 blockade. *Adv. Mater.* **31**, 1805888 (2019).
190. Cheng, L., Zhang, X., Tang, J., Lv, Q. & Liu, J. Gene-engineered exosomes-thermosensitive liposomes hybrid nanovesicles by the blockade of CD47 signal for combined photothermal therapy and cancer immunotherapy. *Biomaterials* **275**, 120964 (2021).
191. Chang, M. et al. Colorectal tumor microenvironment-activated bio-decomposable and metabolizable Cu₂O@CaCO₃ nanocomposites for synergistic oncology. *Adv. Mater.* **32**, 2004647 (2020).
192. Zhang, X. et al. A MXene-based bionic cascaded-enzyme nanoreactor for tumor phototherapy/enzyme dynamic therapy and hypoxia-activated chemotherapy. *Nanomicro Lett.* **14**, 22 (2021).
193. Puccetti, P. & Grohmann, U. IDO and regulatory T cells: a role for reverse signalling and non-canonical NF- κ B activation. *Nat. Rev. Immunol.* **7**, 817–823 (2007).
194. Mellor, A. L. & Munn, D. H. IDO expression by dendritic cells: tolerance and tryptophan catabolism. *Nat. Rev. Immunol.* **4**, 762–774 (2004).
195. Patel, C. H., Leone, R. D., Horton, M. R. & Powell, J. D. Targeting metabolism to regulate immune responses in autoimmunity and cancer. *Nat. Rev. Drug Discov.* **18**, 669–688 (2019).
196. Wang, Y. et al. Immunogenic-cell-killing and immunosuppression-inhibiting nanomedicine. *Bioact. Mater.* **6**, 1513–1527 (2021).
197. Le Naour, J., Galluzzi, L., Zitvogel, L., Kroemer, G. & Vacchelli, E. Trial watch: IDO inhibitors in cancer therapy. *Oncoimmunology* **9**, 177625 (2020).
198. Peng, J. et al. Photosensitizer micelles together with IDO inhibitor enhance cancer photothermal therapy and immunotherapy. *Adv. Sci.* **5**, 1700891 (2018).
199. Guo, Y. X. et al. Indoleamine 2,3-dioxygenase (IDO) inhibitors and their nanomedicines for cancer immunotherapy. *Biomaterials* **276**, 121018 (2021).
200. Jiang, W. et al. Reversing immunosuppression in hypoxic and immune-cold tumors with ultrathin oxygen self-supplementing polymer nanosheets under near infrared light irradiation. *Adv. Funct. Mater.* **31**, 2100354 (2021).
201. Ren, X. et al. Intelligent nanomedicine approaches using medical gas-mediated multi-therapeutic modalities against cancer. *J. Biomed. Nanotechnol.* **18**, 24–49 (2022).
202. Kashiwagi, S. et al. Perivascular nitric oxide gradients normalize tumor vasculature. *Nat. Med.* **14**, 255–257 (2008).
203. Sung, Y. C. et al. Delivery of nitric oxide with a nanocarrier promotes tumour vessel normalization and potentiates anti-cancer therapies. *Nat. Nanotechnol.* **14**, 1160–1169 (2019).
204. Sadelain, M., Rivière, I. & Riddell, S. Therapeutic T cell engineering. *Nature* **545**, 423–431 (2017).
205. Perez, C. R. & De Palma, M. Engineering dendritic cell vaccines to improve cancer immunotherapy. *Nat. Commun.* **10**, 5408 (2019).
206. Weber, E. W., Maus, M. V. & Mackall, C. L. The emerging landscape of immune cell therapies. *Cell* **181**, 46–62 (2020).
207. Santos, P. M. & Butterfield, L. H. Dendritic cell-based cancer vaccines. *J. Immunol.* **200**, 443–449 (2018).
208. Lu, J. & Jiang, G. The journey of CAR-T therapy in hematological malignancies. *Mol. Cancer* **21**, 194 (2022).
209. Zhang, H., Bu, C., Peng, Z., Luo, M. & Li, C. The efficacy and safety of anti-CLL1 based CAR-T cells in children with relapsed or refractory acute myeloid leukemia: a multicenter interim analysis. *J. Clin. Oncol.* **39**, 10000 (2021).
210. Ying, Z. et al. Relmacabtagene autoleucel (relma-cel) CD19 CAR-T therapy for adults with heavily pretreated relapsed/refractory large B-cell lymphoma in China. *Cancer Med.* **10**, 999–1011 (2021).
211. Piehler, S. et al. Hyperthermia affects collagen fiber architecture and induces apoptosis in pancreatic and fibroblast tumor hetero-spheroids in vitro. *Nanomedicine* **28**, 102183 (2020).
212. Beola, L. et al. Dual role of magnetic nanoparticles as intracellular hotspots and extracellular matrix disruptors triggered by magnetic hyperthermia in 3D cell culture models. *ACS Appl. Mater. Interfaces* **10**, 44301–44313 (2018).
213. Gouarderes, S., Mingotaud, A. F., Vicendo, P. & Gibot, L. Vascular and extracellular matrix remodeling by physical approaches to improve drug delivery at the tumor site. *Expert Opin. Drug Deliv.* **17**, 1703–1726 (2020).
214. Chen, Q. et al. Photothermal therapy promotes tumor infiltration and antitumor activity of CAR T cells. *Adv. Mater.* **31**, 1900192 (2019).
215. Ye, Y. et al. A melanin-mediated cancer immunotherapy patch. *Sci. Immunol.* **2**, ea5692 (2017).
216. Bear, A. S. et al. Elimination of metastatic melanoma using gold nanoshell-enabled photothermal therapy and adoptive T cell transfer. *PLoS One* **8**, e69073 (2013).
217. Miller, I. C. et al. Enhanced intratumoral activity of CAR T cells engineered to produce immunomodulators under photothermal control. *Nat. Biomed. Eng.* **5**, 1348–1359 (2021).
218. Xiong, R. et al. Photothermal nanofibers enable safe engineering of therapeutic cells. *Nat. Nanotechnol.* **16**, 1281–1291 (2021).
219. Dranoff, G. Cytokines in cancer pathogenesis and cancer therapy. *Nat. Rev. Cancer* **4**, 11–22 (2004).
220. Briukhovetska, D. et al. Interleukins in cancer: from biology to therapy. *Nat. Rev. Cancer* **21**, 481–499 (2021).
221. Propper, D. J. & Balkwill, F. R. Harnessing cytokines and chemokines for cancer therapy. *Nat. Rev. Clin. Oncol.* **19**, 237–253 (2022).
222. Huttmacher, C. & Neri, D. Antibody-cytokine fusion proteins: Biopharmaceuticals with immunomodulatory properties for cancer therapy. *Adv. Drug Deliv. Rev.* **141**, 67–91 (2019).
223. Huang, H., Feng, W., Chen, Y. & Shi, J. Inorganic nanoparticles in clinical trials and translations. *Nano Today* **35**, 100972 (2020).
224. Liu, Y. et al. Metal-based nanoenhancers for future radiotherapy: radiosensitizing and synergistic effects on tumor cells. *Theranostics* **8**, 1824–1849 (2018).
225. Tamarkin, L., Myer, L., Haynes, R. & Paciotti, G. CYT-6091 (Aurimune™): a colloidal gold-based tumor-targeted nanomedicine. *MRS Proc.* **1019**, FF01–FF10 (2007).
226. Teo, P. et al. Complex of TNF- α and modified Fe₃O₄ nanoparticles suppresses tumor growth by magnetic induction hyperthermia. *Cancer Biother. Radiopharm.* **32**, 379–386 (2017).
227. Lin, X. et al. Localized NIR-II photo-immunotherapy through the combination of photothermal ablation and in situ generated interleukin-12 cytokine for efficiently eliminating primary and abscopal tumors. *Nanoscale* **13**, 1745–1758 (2021).
228. Antonio Chiocca, E. Oncolytic viruses. *Nat. Rev. Cancer* **2**, 938–950 (2002).
229. Kaufman, H. L., Kohlhaup, F. J. & Zloza, A. Oncolytic viruses: a new class of immunotherapy drugs. *Nat. Rev. Drug Discov.* **14**, 642–662 (2015).
230. Tian, Y., Xie, D. & Yang, L. Engineering strategies to enhance oncolytic viruses in cancer immunotherapy. *Signal Transduct. Target. Ther.* **7**, 117 (2022).
231. Dolgin, E. Oncolytic viruses get a boost with first FDA-approval recommendation. *Nat. Rev. Drug Discov.* **14**, 369–371 (2015).
232. Volker, S. A new strategy of cancer immunotherapy combining hyperthermia/oncolytic virus pretreatment with specific autologous anti-tumor vaccination. *Cancer Vaccines* **2**, 1006 (2017).
233. Chen, R. et al. An analytical incorporation for temperature distributions in hepatic radiofrequency ablation incorporating the heat-sink effect of large vessels. *Phys. Med. Biol.* **63**, 235026 (2018).
234. Zhang, X. M. et al. Methotrexate-loaded PLGA nanobubbles for ultrasound imaging and synergistic targeted therapy of residual tumor during HIFU ablation. *Biomaterials* **35**, 5148–5161 (2014).
235. VanOsdol, J. et al. Sequential HIFU heating and nanobubble encapsulation provide efficient drug penetration from stealth and temperature sensitive liposomes in colon cancer. *J. Control. Rel.* **247**, 55–63 (2017).
236. Sweeney, E. E., Cano Mejia, J. & Fernandes, R. Photothermal therapy generates a thermal window of immunogenic cell death in neuroblastoma. *Small* **14**, 1800678 (2018).
237. Nguyen, H. T., Tran, K. K., Sun, B. B. & Shen, H. Activation of inflammasomes by tumor cell death mediated by gold nanoshells. *Biomaterials* **33**, 2197–2205 (2012).
238. Kokuryo, D., Kumamoto, E. & Kuroda, K. Recent technological advancements in thermometry. *Adv. Drug Deliv. Rev.* **163**, 19–39 (2020).
239. Saccomandi, P., Schena, E. & Pacella, C. M. in *Image-guided Laser Ablation* (eds Pacella, C. M., Jiang, T. & Mauri, G.) 145–151 (Springer, 2020).
240. Zhao, T. R., Desjardins, A. E., Ourselin, S., Vercauteren, T. & Xia, W. F. Minimally invasive photoacoustic imaging: current status and future perspectives. *Photoacoustics* **16**, 100146 (2019).
241. Goldenberg, S. L., Nir, G. & Salcudean, S. E. A new era: artificial intelligence and machine learning in prostate cancer. *Nat. Rev. Urol.* **16**, 391–403 (2019).
242. Acosta, J. N., Falcone, G. J., Rajpurkar, P. & Topol, E. J. Multimodal biomedical AI. *Nat. Med.* **28**, 1773–1784 (2022).
243. Denu, R. A. et al. Influence of patient, physician, and hospital characteristics on the receipt of guideline-concordant care for inflammatory breast cancer. *Cancer Epidemiol.* **40**, 7–14 (2016).
244. Zhou, J., Zeng, Z. & Li, L. A meta-analysis of Watson for oncology in clinical application. *Sci. Rep.* **11**, 5792 (2021).
245. Sindhvani, S. et al. The entry of nanoparticles into solid tumours. *Nat. Mater.* **19**, 566–575 (2020).
246. de Lázaro, I. & Mooney, D. J. Obstacles and opportunities in a forward vision for cancer nanomedicine. *Nat. Mater.* **20**, 1469–1479 (2021).
247. Stapleton, S. et al. Radiation and heat improve the delivery and efficacy of nanotherapeutics by modulating intratumoral fluid dynamics. *ACS Nano* **12**, 7583–7600 (2018).
248. Theek, B. et al. Sonoporation enhances liposome accumulation and penetration in tumors with low EPR. *J. Control. Rel.* **231**, 77–85 (2016).
249. Golombek, S. K. et al. Tumor targeting via EPR: strategies to enhance patient responses. *Adv. Drug Deliv. Rev.* **130**, 17–38 (2018).
250. Chauhan, V. P. et al. Normalization of tumour blood vessels improves the delivery of nanomedicines in a size-dependent manner. *Nat. Nanotechnol.* **7**, 383–388 (2012).
251. Tong, R. T. et al. Vascular normalization by vascular endothelial growth factor receptor 2 blockade induces a pressure gradient across the vasculature and improves drug penetration in tumors. *Cancer Res.* **64**, 3731–3736 (2004).
252. Nel, A., Ruuslahti, E. & Meng, H. New insights into “permeability” as in the enhanced permeability and retention effect of cancer nanotherapeutics. *ACS Nano* **11**, 9567–9569 (2017).

253. Autio, K. A. et al. Safety and efficacy of BIND-014, a docetaxel nanoparticle targeting prostate-specific membrane antigen for patients with metastatic castration-resistant prostate cancer a phase 2 clinical trial. *JAMA Oncol.* **4**, 1344–1351 (2018).
254. Wheeler, K. E. et al. Environmental dimensions of the protein corona. *Nat. Nanotechnol.* **16**, 617–629 (2021).
255. Witwer, K. W. & Wolfram, J. Extracellular vesicles versus synthetic nanoparticles for drug delivery. *Nat. Rev. Mater.* **6**, 103–106 (2021).
256. FDA. 501(k) Premarket Notification 042216 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K042216> (2022).
257. FDA. Starburst <https://fda.report/GUDID/H7877001023300> (2018).
258. FDA. 501(k) Summary (K140495) https://www.accessdata.fda.gov/cdrh_docs/pdf14/K140495.pdf (2014).
259. FDA. 501(k) Premarket Notification K073207 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K073207> (2022).
260. FDA. 510(k) Premarket Notification K093855 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K093855> (2022).
261. FDA. 510(k) Premarket Notification K093008 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K093008> (2010).
262. FDA. Letter to Baylis Medical Company Inc re. K161949 https://www.accessdata.fda.gov/cdrh_docs/pdf16/K161949.pdf (2017).
263. Whooley, S. FDA clears Stryker bone tumor ablation system. *Mass Device* <https://www.massdevice.com/fda-clears-stryker-bone-tumor-ablation/> (2022).
264. FDA. Letter to Covidien LLC re. K193232 https://www.accessdata.fda.gov/cdrh_docs/pdf19/K193232.pdf (2020).
265. FDA. 510(k) Premarket Notification K173756 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K173756> (2018).
266. FDA. 510(k) Premarket Notification K083157 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K083157> (2009).
267. Microsulis wins FDA approval for upgraded microwave tissue ablation applicator. *Medical Device Network* <https://www.medicaldevice-network.com/uncategorized/news/microsulis-wins-fda-approval-for-upgraded-microwave-tissue-ablation-applicator/> (2012).
268. FDA clears Covidien Ltd evident microwave ablation system for use in nonresectable liver tumor ablation. *Biospace* <https://www.biospace.com/article/releases/fda-clears-covidien-ltd-evident-microwave-ablation-system-for-use-in-nonresectable-liver-tumor-ablation/> (2008).
269. FDA. 510(k) Summary Visualase Thermal Therapy System https://www.accessdata.fda.gov/cdrh_docs/pdf8/K081656.pdf (2008).
270. FDA. Letter to Monteris Medical re K201056 https://www.accessdata.fda.gov/cdrh_docs/pdf20/K201056.pdf (2020).
271. FDA. 510(k) Premarket Notification K160942 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K160942> (2016).
272. FDA. 510(k) Premarket Notification K153023 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=K153023> (2015).
273. FDA. Letter to Profound Medical Inc re. K191200 https://www.accessdata.fda.gov/cdrh_docs/pdf19/K191200.pdf (2019).
274. FDA. Premarket Approval (PMA) P150038 <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpma/pma.cfm?id=P150038S006> (2018).
275. FDA. Recently approved devices h190003 <https://www.fda.gov/medical-devices/recently-approved-devices/sonalleve-mr-hifu-h190003> (2020).
276. Ritchie, R. W. et al. Extracorporeal high intensity focused ultrasound for renal tumours: a 3-year follow-up. *BJU Int.* **106**, 1004–1009 (2010).
277. Zhao, H. et al. Concurrent gemcitabine and high-intensity focused ultrasound therapy in patients with locally advanced pancreatic cancer. *Anticancer Drugs* **21**, 447–452 (2010).
278. Onik, G. et al. Abstract 6540: Regression of metastatic cancer and abscopal effects following in situ vaccination by cryosurgical tumor cell lysis and intratumoral immunotherapy: a case series. *Cancer Res.* **80**, 6540–6540 (2020).

Acknowledgements

X.Z., G.Y. and M.G. acknowledge support from Harvard/MIT. Z.Y. acknowledges research support from the National Natural Science Foundation of China (51703178), China Postdoctoral Science Foundation (2019M663742), Natural Science Foundation of Shaanxi Province (2022JM183), Natural Science Foundation of Zhejiang Province (LWY20H180002), Shaanxi Provincial Key R&D Program (2022SF-342) and Fundamental Research Funds for the Central Universities (xpt012022030, xtr052022012). D.G. acknowledges research support from the National Natural Science Foundation of China (51903203), China Postdoctoral Science Foundation (2019M653661) and Fundamental Research Funds for the Central Universities (xzy012022037). L.J. acknowledges research support from The Program of Innovative Research Team (in Science and Technology) University of Henan Province (23IRTSTHN008) and the Zhongyuan Thousand Talents Plan. Z.T. acknowledges research support from the National Natural Science Foundation of China (82070751).

Author contributions

Z.Y., D.G., J.Z., G.Y., Y.W., X.R., L.J. and X.Z. researched data for the manuscript and wrote the manuscript. All authors made a substantial contribution to discussions of content and reviewed and/or edited the manuscript prior to submission.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41571-022-00717-y>.

Correspondence should be addressed to Zhe Yang, Lin Jin, Zhongmin Tian or Xingcai Zhang.

Peer review information *Nature Reviews Clinical Oncology* thanks H. Zhang and the other, anonymous, reviewer(s), for their contribution to the peer review of this work.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

© Springer Nature Limited 2023